

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34

3D printing in situ gelification of κ -carrageenan solutions: effect of printing variables on the rheological response

I. Díaz^a, C. Gallegos^b, E. Brito-de la Fuente^b, I. Martínez^{a,c}, C. Valencia^{a,c}, M.C. Sánchez^{a,c}, M.J. Díaz^{a,c}, J.M. Franco^{a,c}, ✉

^a Dept. Ingeniería Química, Universidad de Huelva, Campus de El Carmen, 21071 Huelva, Spain

^b Product & Process Engineering Centre, Fresenius Kabi Deutschland GmbH, 61352 Bad Homburg, Germany

^c Chemical Process and Product Technology Research Centre (Pro2TecS), 21071 Huelva, Spain

✉ Corresponding author:

Prof. J.M. Franco. E-mail: franco@uhu.es

ABSTRACT

This work reports a successful 3D printing-based in situ temperature-induced gelification procedure of κ -carrageenan aqueous dispersions. 3D printer was modified to handle low viscosity fluid feeding and more efficiently distribute ambient air at room temperature causing forced convection to accelerate the cooling of the printed layer. Thus, obtained gel samples, containing **30 mg/g** κ -carrageenan in water, showed self-sustaining capability and a rheological response comparable with a reference conventionally prepared gel. Moreover, the effect of main printing variables, such as temperature of the hotend, printing speed and layer height, on the linear viscoelastic response of the gels was analysed by application of the response surface methodology (RSM). In general, gel strength linearly increases by decreasing printing speed and layer height whereas not noticeable improvement in gel strength was achieved by applying hotend temperatures above 80-85 °C. Based on the results obtained from this analysis, an optimisation method is proposed to minimise the temperature and time needed to 3D print a gel with pre-set rheological properties. Overall, this study demonstrates that it is possible to generate in situ 3D printed gel materials with potential uses in food and pharmaco-nutrition, without the aid of reactive additives or initiators, and using a facile protocol.

Keywords:

3D printing, additive manufacturing, κ -carrageenan, food design, gel, Rheology

35 1. Introduction

36 3D printing is becoming a more important part of our lives every day, apart from already being used
37 industrially, due to the wide variety of possibilities that it can deliver, especially in the development
38 of customised and innovative products. There are several technologies that can be included in the
39 category of 3D printing or additive manufacturing. However, those based on fused filament
40 fabrication are probably the most extended, being suitable for a huge range of applications and
41 materials (Jose, Rodriguez, Dixon, Omenetto, & Kaplan, 2016; Ren, Shao, Lin, & Zheng, 2016;
42 Wegrzyn, Golding, & Archer, 2012). Additive manufacturing refers to the process of building a 3D
43 object by adding layer-upon-layer materials, such as plastic, metal, concrete..., or any material
44 capable of forming stable self-supporting structures. The simplicity of this process is what makes it
45 so versatile, since it does not need the use of materials with special reactive properties and/or
46 initiators. In this context, many industries are focusing their interest on adapting 3D printing
47 technology to the specific needs of each sector. Among many other fields, food processing,
48 gastronomy and cooking are showing growing interest in 3D printing technologies (Lipton, Cutler,
49 Nigl, Cohen, & Lipson, 2015; Pallottino et al., 2016; Sun et al., 2015; Sun, Zhou, Huang, Fuh, & Hong,
50 2015; Sun, Zhou, Yan, Huang, & Lin, 2018). In one hand, it can be used in food industry for the
51 development of new products (Godoi, Prakash, & Bhandari, 2016; Lipton et al., 2015). Moreover, it
52 could also be the next cooking utensil present in our kitchens, as blenders or toasters are nowadays
53 (Izdebska & Zołek-Tryznowska, 2016). Beyond this, 3D printing can become an useful tool for
54 pharmaceutical and clinical nutrition industries, since it could be a way to process fully customisable
55 products, according to the needs of each specific patient (Severini & Derossi, 2016; Sun et al., 2018),
56 **e.g. in the design of personalised foods for dysphagia management.** In addition, the in situ product
57 manufacture, just before administration or use, may solve some problems related to storage of
58 unstable or incompatible components.

59 In all these fields, materials with specific gel characteristics are of great importance. Particularly
60 interesting to form hydrogels is κ -carrageenan, which is considered a representative gel forming
61 polysaccharide, with thermoreversible gelling capacity and widely extended use in many
62 applications, especially in the food, pharmaceutical and cosmetic industries (Ikeda & Nishinari, 2001;
63 Liu, Huang, & Li, 2016). As well-known, κ -carrageenan is a sulphated polysaccharide extracted from
64 different species of red seaweed (Yang, Yang, & Yang, 2017), which needs to be heated and then
65 cooled to gelify, what is known as “cold-setting” (Piculell, 2006). It is well accepted the two-step
66 gelation mechanism involving the formation of helical dimers first, as proposed by Rochas and
67 Rinaudo (Rochas & Rinaudo, 1982), followed by the aggregation of double helices (Iijima, Takahashi,
68 Hatakeyama, & Hatakeyama, 2013; Mangione, Giacomazza, Bulone, Martorana, & San Biagio, 2003).

69 In this aggregation process, the molecular chains extend and associate to other molecules favouring
70 the formation of a three-dimensional network composed of double-helical aggregates as the
71 junctions and flexible chains connecting the junctions (Iijima et al., 2013; Liu, Chan, & Li, 2015). To
72 induce the gelation of κ -carrageenan in water, the temperature must be increased, yielding the
73 hydration and swelling of particles in a range of between 40 and 60 °C, which causes an increase in
74 its viscosity. Afterwards, between 70 and 80 °C, a further drop in viscosity occurs, due to the
75 carrageenan molecules being organised in randomly arranged coils (Imeson, 2009). Typical
76 procedures to prepare κ -carrageenan gels deal with heating at 70-90 °C under stirring for 30 min to
77 2h (Ikeda & Nishinari, 2001; Liu et al., 2015; Yang et al., 2017). When the solution is cooled down,
78 these coils are structured giving rise to the formation of the gel network (Ikeda & Nishinari, 2001;
79 Imeson, 2009). Then, a thermal hysteresis is apparent in subsequent heating and cooling cycles
80 above a critical concentration (Yang et al., 2017), being the melting temperature higher than the
81 gelling temperature. However, as well known, the κ -carrageenan gelification process is highly
82 dependent on salt concentration and nature of the cations (Funami et al., 2007; Hermansson,
83 Eriksson, & Jordansson, 1991; Thrimawithana, Young, Dunstan, & Alany, 2010). The need for all
84 these thermal steps to take place in order to produce gelification complicates the use of continuous
85 processes, as is the case of 3D printing. In addition, the specific conditions under which gelling takes
86 place can lead to structures with different properties (Cavallieri, Fialho, & Cunha, 2011; Cooke et al.,
87 2018). In this sense, rheology is presented as a powerful tool to assess the gel characteristics and
88 final performance as a gel matrix in more complex formulations (Liu et al., 2015). The possibility of
89 preparing gel matrices with tailored rheological properties using efficient, quick and facile protocols
90 is a big challenge that can be achieved by properly adapting the 3D printing technology. For instance,
91 in the case of κ -carrageenan gels, it is worth mentioning that the transition from random coil to
92 double helices is extremely fast (Norton, Goodall, Morris, & Rees, 1983). However, subsequent
93 aggregation of double helices to complete the formation of a gel three dimensional network may
94 take up to 15 h (Tecante & Doublier, 1999).

95 Up to now, the application of 3D printing in gel food processing has been mainly limited to extrude
96 with sophisticated shapes previously gellified systems (Chinga-Carrasco et al., 2018; Holland, Tuck, &
97 Foster, 2018; Wang, Zhang, Bhandari, & Yang, 2018) or, in the best cases, a viscoelastic dispersion or
98 solution (Vancauwenberghe et al., 2017). Only a few works on 3D printing-induced gelation, i.e. the
99 so-called 3D gel printing, have been reported for non-food systems (Moxon et al., 2017; Zhu et al.,
100 2016) and most of them required a catalyst or initiator (Ren et al., 2016). It is apparent that 3D
101 printing-induced gelation could also have a relevant application for designing alternative, alpha-

102 amylase-resistant, customised “fresh” pudding-like products for nutritional support of dysphagia
103 patients (Gallegos, Brito-de la Fuente, Clavé, Costa, & Assegehegn, 2017; Turcanu et al., 2018).
104 Taking into account these considerations, the aim of this study was to take advantage of the
105 potential and versatility of the 3D printing technology to induce the in situ gelification of κ -
106 carrageenan aqueous solutions, subsequently studying, by means of a RSM experimental design, the
107 influence of the main 3D printing variables on the rheological properties of the gel matrices
108 obtained.

109

110 **2. Experimental**

111 **2.1. Materials**

112 κ -carrageenan (M_w 193 - 324 kDa, $K \leq 110$ mg/g, $Ca \leq 35$ mg/g, $Na \leq 20$ mg/g, moisture ≤ 120 mg/g)
113 was used as received from Sigma-Aldrich (Germany). Water was directly taken from the Huelva
114 water supply network, with an average content of cations of Ca 15.7 ± 2 mg/L, Mg 10.5 ± 1.4 mg/L, Na
115 29.9 ± 3.6 mg/L, $K < 2$ mg/L (Ayuntamiento de Huelva, 2015; ENAC, 2017). The content and
116 proportions of cations have not been modified during the study to avoid interferences with the
117 effects caused by the variation of the printing parameters.

118 **2.2. Preparation of κ -carrageenan hydrogels**

119 κ -carrageenan hydrogels studied were prepared by 3D printing. A BQ Hephestos 3D printer DIY kit,
120 with Marlin-derived firmware designed by BQ (Spain), was used for this study once previously
121 adapted and submitted to modifications to print liquid feeds, as described in section 3. All the
122 additional components needed for its adaptation were supplied by 3DEspaña (Spain). 3D printer
123 feeding premix was directly prepared by dispersing 30 mg/g κ -carrageenan in water at room
124 temperature under vigorous mixing (1500 rpm), for 30 min, with a magnetic stirrer, without any
125 other supplement of cations, aiming to reproduce ordinary homemade preparations. These
126 dispersions were fed on demand to the printer through a syringe pump system and gellified in situ
127 during the 3D printing process. Main printing variables (printing speed, hotend temperature and
128 layer height) were modified according to the experimental design described below. Carrageenan gel
129 discs of 50 mm diameter and 1 mm height were printed for rheological characterization. The
130 different shapes and models printed during this study, as well as all accessories required to modify
131 the printer, were designed using AutoCAD 2018 software and Tinkercad online app. The slicing of the
132 3D models, necessary to obtain the .gcode extension files with all the information the printer needs
133 to run, was carried out with Ultimaker Cura software and control of the 3D printer during operation
134 was made possible by Repetier Host software.

135 In addition, a reference hydrogel sample was prepared from the same 30 mg/g carrageenan
136 dispersion employed to feed the printer, using a conventional protocol (Liu et al., 2015, 2016). The
137 carrageenan dispersion was heated by means of a heating plate at 80 °C for 2 h in a closed vessel,
138 under vigorous stirring (1500 rpm) and then cooled down to room temperature by natural
139 convection.

140 **2.3. Rheological characterization**

141 Rheological characterisation of gel samples was carried out with a controlled stress rheometer
142 (Physica MCR-301, Anton Paar, Austria). Small-amplitude oscillatory shear (SAOS) tests were
143 performed inside the linear viscoelastic region, using a plate-plate geometry (25 mm diameter, 1
144 mm gap) in a frequency range of 0.03-100 rad/s, at 25 °C. The 50 mm diameter gel discs directly
145 obtained by 3D printing were placed on the rheometer geometry and the excess sample was
146 carefully removed to suit the 25 mm plate diameter.

147 **2.4. Experimental design and statistical analysis**

148 The main printing parameters that have been considered throughout this study are the hotend
149 temperature (T), printing speed (S) and layer height (H). The temperature of the hotend is the
150 temperature of the heating block, which can be set to the desired value and measured continuously;
151 the printing speed is the speed corresponding to the displacement movement of the hotend during
152 the deposition of the material; and the layer height is the thickness of each of these deposited
153 layers. In order to determine the effect of these printing parameters and the interactions between
154 them on the rheological response of the carrageenan gels, with the minimum possible number of
155 experiments, a response surface methodology (RSM) experimental design was applied. In particular,
156 a Box-Wilson three factors circumscribed central composite design (CCC) at 5 levels of each factor,
157 with an α -value of ± 1.682 . Several replicates of the central point of the cube were carried out, with
158 additional experiments lying at the cube vertices and side centres, originating 20 different case
159 studies, which were rheologically characterized.

160 This experimental design enables the construction of second-order polynomials in the independent
161 variables, as shown below:

$$162 Y = a + a_1x_1 + a_2x_2 + a_3x_3 + a_{12}x_1x_2 + a_{13}x_1x_3 + a_{23}x_2x_3 + a_{11}x_1^2 + a_{22}x_2^2 + a_{33}x_3^2 \quad (1)$$

163 Being Y the dependent variable, x_i the normalised values of the independent variables and a_{ij} the
164 constants obtained from regression analysis.

165 In this way, a regression analysis was carried out on selected rheological parameters (see eq. 4
166 parameters defined below) as dependent variables and the printing parameters (printing speed,
167 hotend temperature and layer height) as independent variables. Those variables whose significance
168 level exceeds 0.05 in Student's t test were excluded from the analysis and the resulting polynomials.

169 The normalisation of the values of each variable, necessary to be able to compare its influence, was
170 carried out by means of the following equation:

$$171 \quad X_N = \frac{2 \cdot (X - X_{med})}{(X_{max} - X_{min})} \quad (2)$$

172 Where X_N is the normalised value of the variable, X is its real value, X_{med} is the average value of the
173 variable and X_{max} and X_{min} are the upper and lower limits, respectively, of the range considered for
174 each variable. The range of each variable studied and their normalised values are detailed in Table 1.

175

176 **3. 3D printer setup**

177 Nowadays, there are many different models of 3D printers available, fully assembled and ready to
178 use, which are marketed by different companies. However, most of them do not allow to either
179 handle fluid feed or to introduce modifications to do it. Moreover, for the moment, their prices can
180 be excessive for private consumers and small businesses or institutions. An alternative that is leading
181 to what may be considered a new industrial revolution is the RepRap 3D printing technology. The
182 RepRap (replicating rapid prototype) project is an initiative dedicated to designing and developing
183 low-cost 3D printers. RepRap 3D printers are based on open-source hardware and software, which
184 are able to print most of their own components, so they can self-replicate and repair themselves,
185 making them freely available for everyone (Wittbrodt et al., 2013). Their design, assembly and
186 operating instructions can be found accessible and fully detailed on the web, as well as their
187 firmware code. Thus, their tuning possibilities are almost unlimited. In fact, RepRap community is
188 now made up of hundreds of collaborators, leading to a continuous open-source research and
189 development activity. Therefore, RepRap appears as a technically and economically viable form of
190 additive layer manufacturing of polymer-based products (Kreiger & Pearce, 2013). Moreover, it has
191 the potential to have a lower environmental impact than conventional manufacturing for a variety of
192 products (Kreiger & Pearce, 2013; Pearce et al., 2010).

193 The aim of this work was to go a step forward using a RepRap 3D printer instead of fully assembled
194 commercial printers, in order to achieve gel structures through direct sol-gel transition, i.e. the in
195 situ gelification. This requires the addition and/or replacement of some components of the original
196 printer setup. The proposed self-adapted setup consists of a syringe pump that, through a
197 polytetrafluoroethylene (PTFE) tube, feeds a watertight hotend that provokes the gelification of the
198 carrageenan solution by temperature. Since feeding syringe is not attached to the printing head, it is
199 possible to avoid problems associated with excessive inertia during movement, resulting in higher
200 printing accuracy. A picture and a schematic depiction of this setup can be seen in Figure 1. The
201 syringe pump consists of a plastic structure, designed and printed with the printer itself prior to
202 modification, metal rods as guides for its movement and a motor connected directly to the printer

203 electronics, instead of the motor corresponding to the original extruder. This extruder, prepared to
204 work with plastic filaments, had to be replaced by a Bowden type watertight hotend in order to
205 handle low viscosity fluids. With these modifications, the printer operated as intended and there is
206 no material leakage. Unfortunately, printing definition was very poor and the material flowed when
207 deposited, so that the model was not able to gain height (Figure 2a). After several attempts under
208 different printing conditions, it was concluded that there was not enough time between deposition
209 of the different layers for cooling and structuring of the material, even when the printing speed was
210 greatly reduced. The Bowden-type hotend does not allow to couple a standard layer fan, so there
211 was no cooling system for the printed material apart from exposure to ambient air. Aiming to solve
212 this drawback, a new piece was designed and printed on polylactic acid (PLA). This piece allows the
213 layer fan of the old extruder to be adapted to the new hotend, as well as distributing the air flow
214 evenly thanks to a diffuser ring (see scheme in Figure 2b). This simple cooling system is based on
215 ambient air at room temperature; but the mere fact of causing forced convection is sufficient to
216 accelerate the cooling of the material, allowing the gel to structure in a way that the model can gain
217 height. These modifications resulted in the printing of carrageenan gel models with an acceptable
218 resolution and self-sustaining capability, as shown in Figure 2c (see also movie placed in the
219 Supporting Information). **However, the resolution in this case is limited by the nature of the material.**
220 **When deposited, the material flows until it cools down sufficiently to structure itself. This is the**
221 **reason why it was not possible to increase the height of the printed gels before the modification of**
222 **the system with the cooling ring. In the same way, this brief flow of material means that the shapes**
223 **are not preserved as faithfully as when working with other more conventional solid materials.**
224 **Dimensions of the star-shaped model printed after optimising the resolution are displayed in Figure**
225 **2c.**

226 Due to these changes on the printer setup, the device had to be recalibrated, making some changes
227 to the printer firmware for proper operation. Since the default extruder stepper motor was replaced
228 by the one that drives the syringe pump, it was necessary to modify the value of axis steps per unit
229 so that the feed flow was adequate for the geometry of the new feed system. **From the values of**
230 **printing speed, layer height and nozzle diameter (the same 0.4 mm diameter nozzle has been used**
231 **for the entire study), the printer automatically adjusts the required material flow.** The firmware was
232 configured to give the correct reading of the new hotend thermocouple and the PID control
233 constants for the new heater block were also determined.

234

235 **4. Results and discussion**

236 **4.1. Rheological behaviour**

237 κ -carrageenan gels processed by 3D printing exhibited a qualitatively similar rheological response,
238 characterized by a slight frequency dependence of the storage modulus (G') and a soft minimum in
239 the evolution of the loss modulus (G''), being G' values almost one decade higher than G'' values.
240 Figure 3 shows the evolution of both SAOS functions with frequency for selected 3D printed
241 carrageenan gels. In particular, Figure 3a presents frequency sweeps of several samples, processed
242 at the same speed (35 mm/s) and layer height (0.18 mm) but applying different temperatures at the
243 hotend, compared with a carrageenan gel conventionally prepared at 80 °C. As can be seen, only the
244 sample printed at 50 °C significantly differs from the others. As well known, at such a low
245 temperature, the necessary mobility of the carrageenan coils is not reached in the solution phase
246 and the gel network structure cannot fully develop when it was subsequently cooled. Once the
247 temperature threshold required for the release of the coils has been exceeded, the structures
248 obtained after gelling are quite similar to each other, even if the hotend temperature is significantly
249 increased, i.e. 110 °C. Above this temperature threshold, both 3D printed and conventionally
250 prepared gel samples have a similar rheological response. In fact, the linear viscoelastic functions of
251 the carrageenan prepared in a conventional heating device, for 2 h under mixing, and those
252 corresponding to the sample obtained by 3D printing at 80 °C are practically identical. Figure 3b, on
253 the other hand, illustrates the effect of layer height at the same temperature (80 °C) and printing
254 speed (35 mm/s), while, finally, Figure 3c compares the SAOS functions of printed gels at different
255 speeds, keeping constant both hotend temperature (80 °C) and layer height (0.18 mm). As can be
256 clearly seen, both higher layer height (H) and higher speed (S) lower the viscoelastic moduli over the
257 entire frequency range considered.

258 The evolution of SAOS functions with frequency obtained in all cases shows a slight decrease in G'
259 when the frequency is reduced, due to the structural rearrangement and relaxation processes
260 undergone by the gel network when it is subjected to a high enough stress. Besides this, moderate
261 differences between G' and G'' values (less than one decade; in most cases, $\tan \delta$ values higher than
262 0.1) is typically reported to be characteristic of relatively “weak gels” (Ross-Murphy & Shatwell,
263 1993). For all the printed gels, the loss tangent exhibited very similar values ($\tan \delta=0.20\pm 0.12$) within
264 the whole frequency range studied. For this reason, the complex modulus (G^*), defined as:

$$265 \quad |G^*|^2 = |G'|^2 + |G''|^2 \quad (3)$$

266 can be selected as an appropriate viscoelastic function to be modelled as a function of the 3D
267 printing variables. G^* has been fitted to a power-law model:

$$268 \quad G^* = A\omega^n \quad (4)$$

269 where ω is the angular frequency (rad/s) and, according to the basic theory of gels, A represents the
270 gel strength and n is a power-law relaxation exponent (Campo-Deaño & Tovar, 2009; Moresi, Bruno,

271 & Parente, 2004; Munarin et al., 2014). In physical gelation, it has been widely reported that physical
272 cross-links, i.e. entanglements, may change with time under the application of stress, causing an
273 actual fluctuation in the state of gelation. For these so-called weak gels, Mita and Bohlin (Mita &
274 Bohlin, 1983) proposed that the real structure of such materials may be represented as a
275 cooperative arrangement of flow units. In this context, the reciprocal of the above referred power
276 law index, $z=1/n$, has been defined by different authors as a coordination number (Gabriele, De
277 Cindio, & D'Antona, 2001) that assumes the meaning of the number of rheological units correlated
278 with one other in the three dimensional structure, and applied to different gel systems (Antunes et
279 al., 2008; Basu et al., 2011; Lupi et al., 2015). Therefore, in the present study, the coordination
280 number could be related to the number of interacting rheological units within the 3D gel network,
281 whilst A is a parameter associated to the strength of the interactions between them and therefore
282 may be correlated to the bulk gel strength. An example of fitting of the G^* vs. frequency plots to eq.
283 (4) can be seen in Fig. S1, in the "Supporting Information" document.

284

285 **4.2. Response Surface modelling**

286 Table 2 lists the values of fitting parameters A and z obtained for each case, as well as the
287 normalised values of independent variables. By applying the multivariate regression analysis,
288 parameter A can be described as a function of the independent variables by means of the following
289 equation:

$$290 \quad A = 11224.0 - 1931.0 \cdot S + 3617.2 \cdot T - 2881.0 \cdot H - 1355.1 \cdot T^2 \quad (5)$$

$$(R^2 = 0.91; F = 37.33; df = 4.15)$$

291 This means that A linearly decreases with the printing speed (S) and layer height (H), as Fig. 3b and
292 3c suggest, whereas it depends quadratically on the temperature (T), in the range of printing
293 variables studied. Moreover, **good fitting results were** obtained without the need to include terms of
294 interaction among variables.

295 A similar dependence of the printing variables was found on parameter z, which can be modelled as
296 follows:

$$297 \quad z = 16.48 - 0.84 \cdot S + 1.11 \cdot T - 1.02 \cdot H - 1.82 \cdot T^2 \quad (6)$$

$$(R^2 = 0.93; F = 47.01; df = 4.15)$$

298 Since independent variables can only be associated with two of the axes of the 3D graphic (the third
299 axis corresponds to the observed dependent variable), the response surfaces are drawn by giving
300 values between -1.682 and +1.682 to two of the independent variables, while the third remains
301 constant (see Figure 4).

302 **4.2.1. Constant layer height (H) plots**

303 As can be deduced from eqs. 5 and 6, A and z are linearly related to H, so that the surfaces
304 corresponding to the different levels of H are parallel and equidistant from each other, as it is shown
305 in Figs. 4a and 4b. As can be seen, parameter A decreases by increasing the layer height. An increase
306 in layer height implies a larger amount of deposited material, so more time is required to cool down
307 a given volume of sample before the next layer is deposited. Therefore, especially at high printing
308 speed, the material does not have enough time to be properly cooled down and the network that is
309 formed exhibits weaker interactions. As a result, the higher the layer height, the lower values of A
310 and z were obtained, which in practice means lower gel strength.

311 For the same H-value, response surface model predicts a tendency to reach maximum A-values for
312 low speeds and high temperatures, which corresponds to the expected behaviour, since it is the
313 combination under which the material is most exposed to temperature increment, due to the longer
314 residence time and the greater temperature gradient between the fed material and the hotend. As
315 the coil-helix transition temperature is more widely exceeded, the carrageenan chains are more
316 mobile and the creation of the gel structure is favoured upon cooling. However, the values of
317 parameter A are levelled off at around 100 °C, whereas z reaches a maximum at around 80-85 °C.
318 This means that no benefit can be taken by excessively heating the sample; on the contrary, a not so
319 compact gel structure seems to be formed due to a partial non-reversible coil-helix transition and/or
320 not so efficient aggregation of double helices. As Tanaka (Tanaka, 2003) pointed out, different
321 associations of double helices, differing in the number of helices and average helix length, may be
322 achieved at different temperatures. In particular, this theoretical study predicts that the number of
323 helices per chain, which may be related with parameter z in eq. (4), passes through a maximum at a
324 given temperature. In addition, low velocities also allow each layer of deposited material more time
325 to cool down before it is covered by the next layer of hot material. So, the optimum point will be
326 determined by the maximum time (minimum speed) that can be used in printing, ensuring that the
327 limit value that allows the printer to work without clogging problems is reached. **In this case, as a
328 water-based material, conditions combining an excessively low flow rate with high temperature (e.g.
329 printing speed below 2 mm/s and temperature above 85 °C) may result in drying of the material and
330 consequent system clogging problems. However, in most applications, the aim is to reduce total
331 printing time and energy consumption, so that operating conditions would be closer to the upper
332 limit: high layer height and printing speed at the lowest possible temperatures.**

333 When hotend temperature is increased enough to allow the dispersion to exceed the hydration and
334 dissolution threshold of carrageenan molecules (above 62-64 °C, see Figure S2 in the Supporting
335 Information), it is not necessary for the material to remain exposed to that temperature for so long,
336 so that printing can be accelerated without major changes in the resulting rheological properties.

337 4.2.2. *Constant printing speed (S) plots*

338 As can be seen in Figs. 4c and 4d, the A and z response surfaces for constant S-values are very similar
339 to those of constant H-values, since the relationship of both independent variables with the
340 dependent variables are linear and comparable (eqs. 5 and 6). Both the layer height and printing
341 speed are related to the material flow, which is being heated inside the hotend, since H determines
342 the amount of material that goes through the 3D printing system and S is related to the hotend
343 residence time. Moreover, once the material has been deposited on printing surface or on the
344 previous layer, there is also an equivalent effect during cooling. The printing speed determines the
345 time the material is cooled and therefore the time during which the material can be properly self-
346 structured, whereas the layer height is related to the amount of material that needs to be self-
347 structured.

348 4.2.3. *Constant temperature (T) plots*

349 Unlike those corresponding to constant H and S values, all graphs constructed for constant T-values
350 are flat surfaces (Figs. 4e and 4f). Moreover, there are no interactions between the variables, so that
351 all surfaces are parallel to each other. As just mentioned, the effects of layer height and print speed
352 are somewhat equivalent. Therefore, the maximum values of A (stronger gel network) and z (higher
353 number on interacting units) are predicted for the lowest values of H and S, while the minimum is
354 exactly at the opposite corner, which tends to zero in the case of A, i.e. extremely low gel
355 consistency. On the other hand, as previously discussed, A values are closer for hotend temperatures
356 around 80-100 °C, and z reaches maximum values at around 80 °C (the higher plane in Fig. 4f).

357 4.3. *Time as a variable*

358 One of the goals of this study was to elaborate a practical model that allows predicting the
359 rheological response of κ -carrageenan gels as a function of the main 3D printing variables directly
360 involved in its manufacture: layer height, printing speed and temperature of the 3D printer hotend.
361 In a typical product design process, the properties to be achieved would be set and the processing
362 conditions should be established to achieve them at the lowest possible cost. In this context, the
363 temperature would be an easily optimisable variable according to energy consumption: the most
364 appropriate value would be the lowest value able to guarantee the desired final properties. For
365 instance, as previously discussed, not noticeable improvement in gel strength was achieved by
366 applying hotend temperatures above 80-85 °C. However, both layer height and printing speed are
367 more complex to analyse. These variables are necessary to be able to define the conditions of the
368 printing process, but do not have as much effect on the cost of the process as in the case of
369 temperature. Nevertheless, the combination of both of them determines the value of a decisive
370 variable in terms of cost optimization: the printing time.

371 Both the printer firmware and the software used in this study are the same as when printing with a
 372 conventional solid filament. In this way, the system can be treated analogously to a traditional solid
 373 filament printing system.

374 The printing speed, S , is the linear speed of the nozzle when printing. Thus:

$$375 \quad t = \frac{L}{S} \quad (7)$$

376 where t is the total printing time and L is the length of the complete path the nozzle has to take to
 377 print a certain model. For a constant section 3D printed object, like the discs printed in this work for
 378 further rheological assessment of gel models:

$$379 \quad L = L_c \cdot N_c \quad (8)$$

380 L_c being the length of the path necessary to print each layer and N_c being the number of layers of
 381 the model. By combining both equations (eqs. 7 and 8):

$$382 \quad t = \frac{L_c \cdot N_c}{S} \quad (9)$$

383 The number of layers for a certain object would be the total height (h_m) divided by the height of each
 384 layer (H). So, the total printing time will be:

$$385 \quad t = \frac{L_c \cdot h_m}{S \cdot H} \quad (10)$$

386 In this equation, h_m is a known value (1 mm in the gel discs) and L_c can be calculated from the
 387 “filament” length, L_{filament} . When the printer is operated conventionally by using a solid plastic
 388 filament that melts before being deposited, slicing software provides the length of filament to be
 389 used for printing a certain model. This value, included in the .gcode file, is the L_{filament} .

390 This filament is of a known diameter (d_{filament}) and, when it melts and passes through the nozzle, its
 391 section changes, taking the width of the outlet hole, d_{nozzle} , and the thickness corresponding to the
 392 layer height, H . If the volume of the material is considered to be constant before and after the nozzle
 393 (regardless of thermal expansion), the length of the filament consumed and the length of the
 394 deposited thread will be different. In this case, as it is not a solid filament, the internal diameter of
 395 the feed tube (2 mm) is taken as d_{filament} .

396 Thus, known d_{filament} and d_{nozzle} , the L_{filament} value, taken from the .gcode file by analogy with a solid
 397 filament, can be used to calculate the length of the deposited thread. Moreover, if the value of
 398 L_{filament} is taken as the data corresponding to laminate a single layer of the discs, the value of L_c can
 399 be obtained as follows:

$$400 \quad L_c \cdot d_{\text{nozzle}} \cdot H = L_{\text{filament}} \cdot \frac{\pi}{4} \cdot (d_{\text{filament}})^2 \quad (11)$$

$$401 \quad L_c = \frac{L_{\text{filament}} \cdot \frac{\pi}{4} \cdot d_{\text{filament}}}{d_{\text{nozzle}} \cdot H} = \frac{L_{\text{filament}} \cdot \frac{\pi}{4} \cdot (2 \text{ mm})^2}{0.4 \text{ mm} \cdot H} = 986.4 \text{ mm} \quad (12)$$

402 When considering discs printed with different H values, L_{filament} also changes (the thicker the
403 deposited thread, the more filament is consumed). Therefore, the path to be followed by the hotend
404 to create a layer (L_c) is always the same, since the surface to be covered and the diameter of the
405 nozzle are also the same. This value is provided by L_{filament} and H-values, corresponding for each disc,
406 and for all of them the same L_c value is obtained (d_{filament} and d_{nozzle} are fixed values). So, for the case
407 of this study:

$$408 \quad t = \frac{L_c \cdot h_m}{S \cdot H} = \frac{986.4 \text{ mm} \cdot 1 \text{ mm}}{S \cdot H} = \frac{986.4 \text{ mm}^2}{S \cdot H} \quad (13)$$

409 The following procedure is an example of the utility of this relationship when, for instance, an A-
410 value is imposed. Starting from the equation obtained to predict the behaviour of parameter A (eq.
411 5) and rearranging:

$$412 \quad S = \frac{-A + 11224.0 + 3617.2 \cdot T - 2881.0 \cdot H - 1355.1 \cdot T^2}{2710.2} \quad (14)$$

413 Thus, for each pair of given H-T values and for a certain fixed value of A, S can be calculated.
414 Subsequently, by substituting each calculated S-value and their corresponding H-values in the eq. 13,
415 the printing time, t, is obtained.

416 It is important at this point to notice that, in eq. 13, the real values of variables S and H are entered,
417 while, in eq. 14, it is necessary to use the normalized values of H and T between -1.682 and +1.682.
418 Likewise, the printing speed values calculated from this equation also correspond to the normalised
419 scale. Also, it is worth mentioning that fixed constant levels of H have been considered in this study,
420 instead of S-values, because the layer height determines the resolution of the final 3D printed
421 product and could, in some cases, be of interest for the optimisation of the printing process.
422 However, when working with this kind of viscoelastic materials and in applications where the
423 resolution of the final 3D object is not decisive, S instead of H may be chosen, following then the
424 same procedure. Thus, for a given value of A (defined by product specifications), a new table can be
425 elaborated from eqs. 13 and 14 (see selected values in Table S1, in the "Supporting Information"
426 document), which in this case has been calculated for $A = 12000 \text{ Pa} \cdot \text{s}^{1/2}$. Using these values, different
427 curves can be plotted (Fig. 5), from which the time and temperature required to obtain a gel with an
428 imposed A-value can be determined. If there would be any restriction to raise the temperature
429 (degradation of some component, for example), by drawing a horizontal line from that value, the
430 first crossing with any of the plotted curves will indicate the minimum printing time that will be used
431 in the process to achieve a target rheological parameter, a value of $A = 12000 \text{ Pa} \cdot \text{s}^{1/2}$ in this example.
432 On the contrary, if it is time that determines, for example, whether it is profitable to print a certain
433 product in 3D, the procedure would be the inverse: it could be determined what would be the
434 minimum temperature value that could be used and, therefore, the associated energy consumption.

435

436 **5. Concluding remarks**

437 3D printed κ -carrageenan gels were successfully obtained by means of a RepRap 3D printer, which
438 was modified to be able to in situ gellify carrageenan-in-water dispersions. Thus printed gels can be
439 obtained in a few minutes, exhibiting a rheological response comparable to that prepared
440 conventionally by a process of 2 h duration. This seems to mean that, in spite of how briefly the
441 material is inside the 3D printer, it is sufficient for the different transitions necessary for the
442 development of the gel network.

443 Moreover, the multivariate analysis carried out on SAOS parameters obtained for 3D printed κ -
444 carrageenan allowed to determine the effect of the main printing parameters (layer height, printing
445 speed and hotend temperature) on the rheological response of gel structures created during printing
446 process. In general, it can be concluded that an elevated temperature, together with a reduced
447 velocity and layer height, favours the structuring of carrageenan, giving rise to gel networks with
448 more intense interactions.

449 Based on this data and taking into account the influence of speed and layer height on printing time,
450 an example of optimization of printing variables for obtaining a final product with a given
451 specification has been presented.

452

453 **6. References**

- 454 Antunes, F. E., Coppola, L., Rossi, C. O., & Ranieri, G. A. (2008). Gelation of charged bio-
455 nanocompartments induced by associative and non-associative polysaccharides. *Colloids and*
456 *Surfaces B: Biointerfaces*, 66(1), 134–140. <https://doi.org/10.1016/j.colsurfb.2008.06.004>
- 457 **Ayuntamiento de Huelva. (2015). Agua potable y parámetros de calidad del agua semanal. Retrieved**
458 **June 22, 2018, from <http://www.aguashuelva.com/ESP/191.asp>**
- 459 Basu, S., Shivhare, U. S., Singh, T. V., & Beniwal, V. S. (2011). Rheological, textural and spectral
460 characteristics of sorbitol substituted mango jam. *Journal of Food Engineering*, 105(3), 503–
461 512. <https://doi.org/10.1016/j.jfoodeng.2011.03.014>
- 462 Campo-Deaño, L., & Tovar, C. (2009, October 1). The effect of egg albumen on the viscoelasticity of
463 crab sticks made from Alaska Pollock and Pacific Whiting surimi. *Food Hydrocolloids*. Elsevier.
464 <https://doi.org/10.1016/j.foodhyd.2009.03.013>
- 465 Cavallieri, A. L. F., Fialho, N. A. V., & Cunha, R. L. (2011). Sodium caseinate and κ -carrageenan
466 interactions in acid gels: Effect of polysaccharide dissolution temperature and sucrose addition.
467 *International Journal of Food Properties*, 14(2), 251–263.
468 <https://doi.org/10.1080/10942910903176345>

469 Chinga-Carrasco, G., Ehman, N. V., Pettersson, J., Vallejos, M., Brodin, M., Felissia, F. E., ... Area, M. C.
470 (2018). Pulping and pretreatment affect the characteristics of bagasse inks for 3D printing. *ACS*
471 *Sustainable Chemistry & Engineering*, 6(3), 4068–4075.
472 <https://doi.org/10.1021/acssuschemeng.7b04440>

473 Cooke, M. E., Jones, S. W., ter Horst, B., Moiemmen, N., Snow, M., Chouhan, G., ... Grover, L. M.
474 (2018). Structuring of Hydrogels across Multiple Length Scales for Biomedical Applications.
475 *Advanced Materials*, 30(14), 1705013. <https://doi.org/10.1002/adma.201705013>

476 ENAC. (2017). *Informe de análisis*. Retrieved from [http://www.aguashuelva.com/DOC/01-02-](http://www.aguashuelva.com/DOC/01-02-2018_Salida_Depsito_N1_2206179.pdf)
477 [2018_Salida_Depsito_N1_2206179.pdf](http://www.aguashuelva.com/DOC/01-02-2018_Salida_Depsito_N1_2206179.pdf)

478 Funami, T., Hiroe, M., Noda, S., Asai, I., Ikeda, S., & Nishinari, K. (2007). Influence of molecular
479 structure imaged with atomic force microscopy on the rheological behavior of carrageenan
480 aqueous systems in the presence or absence of cations. *Food Hydrocolloids*, 21(4), 617–629.
481 <https://doi.org/10.1016/j.foodhyd.2006.07.013>

482 Gabriele, D., De Cindio, B., & D'Antona, P. (2001). A weak gel model for foods. *Rheologica Acta*,
483 40(2), 120–127. <https://doi.org/10.1007/s003970000139>

484 Gallegos, C., Brito-de la Fuente, E., Clavé, P., Costa, A., & Assegehegn, G. (2017). Nutritional Aspects
485 of Dysphagia Management. *Advances in Food and Nutrition Research*, 81, 271–318.
486 <https://doi.org/10.1016/BS.AFNR.2016.11.008>

487 Godoi, F. C., Prakash, S., & Bhandari, B. R. (2016, June 1). 3d printing technologies applied for food
488 design: Status and prospects. *Journal of Food Engineering*. Elsevier.
489 <https://doi.org/10.1016/j.jfoodeng.2016.01.025>

490 Hermansson, A. M., Eriksson, E., & Jordansson, E. (1991). Effects of potassium, sodium and calcium
491 on the microstructure and rheological behaviour of kappa-carrageenan gels. *Carbohydrate*
492 *Polymers*, 16(3), 297–320. [https://doi.org/10.1016/0144-8617\(91\)90115-S](https://doi.org/10.1016/0144-8617(91)90115-S)

493 Holland, S., Tuck, C., & Foster, T. (2018). Fluid Gels: a New Feedstock for High Viscosity Jetting. *Food*
494 *Biophysics*, 13(2), 175–185. <https://doi.org/10.1007/s11483-018-9523-x>

495 Iijima, M., Takahashi, M., Hatakeyama, T., & Hatakeyama, H. (2013). Detailed investigation of gel-sol
496 transition temperature of κ -carrageenan studied by DSC, TMA and FBM. *Journal of Thermal*
497 *Analysis and Calorimetry*, 114(2), 895–901. <https://doi.org/10.1007/s10973-013-2953-2>

498 Ikeda, S., & Nishinari, K. (2001). “Weak gel”-type rheological properties of aqueous dispersions of
499 nonaggregated κ -carrageenan helices. *Journal of Agricultural and Food Chemistry*, 49(9), 4436–
500 4441. <https://doi.org/10.1021/jf0103065>

501 Imeson, A. P. (2009). Carrageenan and furcellaran. In G. O. Phillips & P. A. Williams (Eds.), *Handbook*
502 *of Hydrocolloids: Second Edition* (pp. 164–185). Boca Raton: Elsevier.

503 <https://doi.org/10.1533/9781845695873.164>

504 Izdebska, J., & Zołek-Tryznowska, Z. (2016). 3D food printing - Facts and future. *Agro Food Industry*

505 *Hi-Tech*, 27(2), 33–37. Retrieved from

506 http://www.teknoscienze.com/Contents/Riviste/PDF/AF2_2016_low_35-40.pdf

507 Jose, R. R., Rodriguez, M. J., Dixon, T. A., Omenetto, F., & Kaplan, D. L. (2016). Evolution of Bioinks

508 and Additive Manufacturing Technologies for 3D Bioprinting. *ACS Biomaterials Science and*

509 *Engineering*, 2(10), 1662–1678. <https://doi.org/10.1021/acsbiomaterials.6b00088>

510 Kreiger, M., & Pearce, J. M. (2013). Environmental life cycle analysis of distributed three-dimensional

511 printing and conventional manufacturing of polymer products. *ACS Sustainable Chemistry and*

512 *Engineering*, 1(12), 1511–1519. <https://doi.org/10.1021/sc400093k>

513 Lipton, J. I., Cutler, M., Nigl, F., Cohen, D., & Lipson, H. (2015). Additive manufacturing for the food

514 industry. *Trends in Food Science and Technology*, 43(1), 114–123.

515 <https://doi.org/10.1016/j.tifs.2015.02.004>

516 Liu, S., Chan, W. L., & Li, L. (2015). Rheological Properties and Scaling Laws of κ -Carrageenan in

517 Aqueous Solution. *Macromolecules*, 48(20), 7649–7657.

518 <https://doi.org/10.1021/acs.macromol.5b01922>

519 Liu, S., Huang, S., & Li, L. (2016). Thermoreversible gelation and viscoelasticity of κ -carrageenan

520 hydrogels. *Journal of Rheology*, 60(2), 203–214. <https://doi.org/10.1122/1.4938525>

521 Lupi, F. R., Gentile, L., Gabriele, D., Mazzulla, S., Baldino, N., & de Cindio, B. (2015). Olive oil and

522 hyperthermal water bigels for cosmetic uses. *Journal of Colloid and Interface Science*, 459, 70–

523 78. <https://doi.org/10.1016/j.jcis.2015.08.013>

524 Mangione, M. R., Giacomazza, D., Bulone, D., Martorana, V., & San Biagio, P. L. (2003).

525 Thermoreversible gelation of κ -Carrageenan: Relation between conformational transition and

526 aggregation. *Biophysical Chemistry*, 104(1), 95–105. [https://doi.org/10.1016/S0301-](https://doi.org/10.1016/S0301-4622(02)00341-1)

527 [4622\(02\)00341-1](https://doi.org/10.1016/S0301-4622(02)00341-1)

528 Mita, T., & Bohlin, L. (1983). Shear stress relaxation of chemically modified gluten. *Cereal Chemistry*,

529 60(2), 93–97. Retrieved from <http://ci.nii.ac.jp/naid/80001616483/en/>

530 Moresi, M., Bruno, M., & Parente, E. (2004). Viscoelastic properties of microbial alginate gels by

531 oscillatory dynamic tests. *Journal of Food Engineering*, 64(2), 179–186.

532 <https://doi.org/10.1016/j.jfoodeng.2003.09.030>

533 Moxon, S. R., Cooke, M. E., Cox, S. C., Snow, M., Jeys, L., Jones, S. W., ... Grover, L. M. (2017).

534 *Suspended Manufacture of Biological Structures. Advanced Materials*, 29(13), 1605594.

535 <https://doi.org/10.1002/adma.201605594>

536 Munarin, F., Petrini, P., Barcellona, G., Roversi, T., Piazza, L., Visai, L., & Tanzi, M. C. (2014). Reactive

537 hydroxyapatite fillers for pectin biocomposites. *Materials Science and Engineering: C*, 45, 154–
538 161. <https://doi.org/10.1016/J.MSEC.2014.09.003>

539 Norton, I. T., Goodall, D. M., Morris, E. R., & Rees, D. A. (1983). Equilibrium and dynamic studies of
540 the disorder-order transition of kappa carrageenan. *Journal of the Chemical Society, Faraday*
541 *Transactions 1: Physical Chemistry in Condensed Phases*, 79(10), 2489–2500.
542 <https://doi.org/10.1039/F19837902489>

543 Pallottino, F., Hakola, L., Costa, C., Antonucci, F., Figorilli, S., Seisto, A., & Menesatti, P. (2016).
544 Printing on Food or Food Printing: a Review. *Food and Bioprocess Technology*, 9(5), 725–733.
545 <https://doi.org/10.1007/s11947-016-1692-3>

546 Pearce, J. M., Morris Blair, C., Laciak, K. J., Andrews, R., Nosrat, A., & Zelenika-Zovko, I. (2010). 3-D
547 Printing of Open Source Appropriate Technologies for Self-Directed Sustainable Development.
548 *Journal of Sustainable Development*, 3(4), 17–29. <https://doi.org/10.5539/jsd.v3n4p17>

549 Piculell, L. (2006). Gelling carrageenans. In A. M. Stephen, G. O. Phillips, & P. A. Williams (Eds.), *Food*
550 *Polysaccharides and Their Applications* (2nd ed., pp. 239–287). Boca Raton: CRC Press.

551 Ren, X., Shao, H., Lin, T., & Zheng, H. (2016). 3D gel-printing-An additive manufacturing method for
552 producing complex shape parts. *Materials and Design*, 101, 80–87.
553 <https://doi.org/10.1016/j.matdes.2016.03.152>

554 Rochas, C., & Rinaudo, M. (1982). Calorimetric determination of the conformational transition of
555 kappa carrageenan. *Carbohydrate Research*, 105(2), 227–236. [https://doi.org/10.1016/S0008-6215\(00\)84970-8](https://doi.org/10.1016/S0008-6215(00)84970-8)

557 Ross-Murphy, S. B., & Shatwell, K. P. (1993). Polysaccharide strong and weak gels. *Biorheology*, 30(3–
558 4), 217–227. Retrieved from <http://www.ncbi.nlm.nih.gov/pubmed/8286724>

559 Severini, C., & Derossi, A. (2016). Could the 3D Printing Technology be a Useful Strategy to Obtain
560 Customized Nutrition? *Journal of Clinical Gastroenterology*, 50(December), S175–S178.
561 <https://doi.org/10.1097/MCG.0000000000000705>

562 Sun, J., Peng, Z., Zhou, W., Fuh, J. Y. H., Hong, G. S., & Chiu, A. (2015). A Review on 3D Printing for
563 Customized Food Fabrication. *Procedia Manufacturing*, 1, 308–319.
564 <https://doi.org/10.1016/j.promfg.2015.09.057>

565 Sun, J., Zhou, W., Huang, D., Fuh, J. Y. H., & Hong, G. S. (2015). An Overview of 3D Printing
566 Technologies for Food Fabrication. *Food and Bioprocess Technology*, 8(8), 1605–1615.
567 <https://doi.org/10.1007/s11947-015-1528-6>

568 Sun, J., Zhou, W., Yan, L., Huang, D., & Lin, L. ya. (2018). Extrusion-based food printing for digitalized
569 food design and nutrition control. *Journal of Food Engineering*, 220, 1–11.
570 <https://doi.org/10.1016/j.jfoodeng.2017.02.028>

571 Tanaka, F. (2003). Thermoreversible Gelation Driven by Coil-to-Helix Transition of Polymers.
572 *Macromolecules*, 36(14), 5392–5405. <https://doi.org/10.1021/ma021688y>

573 Tecante, A., & Doublier, J. L. (1999). Steady flow and viscoelastic behavior of crosslinked waxy corn
574 starch- κ -carrageenan pastes and gels. *Carbohydrate Polymers*, 40(3), 221–231.
575 [https://doi.org/10.1016/S0144-8617\(99\)00057-0](https://doi.org/10.1016/S0144-8617(99)00057-0)

576 Thrimawithana, T. R., Young, S., Dunstan, D. E., & Alany, R. G. (2010). Texture and rheological
577 characterization of kappa and iota carrageenan in the presence of counter ions. *Carbohydrate*
578 *Polymers*, 82(1), 69–77. <https://doi.org/10.1016/j.carbpol.2010.04.024>

579 Turcanu, M., Siegert, N., Secouard, S., Brito-de la Fuente, E., Balan, C., & Gallegos, C. (2018). An
580 alternative elongational method to study the effect of saliva on thickened fluids for dysphagia
581 nutritional support. *Journal of Food Engineering*, 228, 79–83.
582 <https://doi.org/10.1016/J.JFOODENG.2018.02.015>

583 Vancauwenberghe, V., Katalagarianakis, L., Wang, Z., Meerts, M., Hertog, M., Verboven, P., ...
584 Nicolai, B. (2017). Pectin based food-ink formulations for 3-D printing of customizable porous
585 food simulants. *Innovative Food Science and Emerging Technologies*, 42, 138–150.
586 <https://doi.org/10.1016/j.ifset.2017.06.011>

587 Wang, L., Zhang, M., Bhandari, B., & Yang, C. (2018). Investigation on fish surimi gel as promising
588 food material for 3D printing. *Journal of Food Engineering*, 220, 101–108.
589 <https://doi.org/10.1016/j.jfoodeng.2017.02.029>

590 Wegrzyn, T. F., Golding, M., & Archer, R. H. (2012). Food Layered Manufacture: A new process for
591 constructing solid foods. *Trends in Food Science and Technology*, 27(2), 66–72.
592 <https://doi.org/10.1016/j.tifs.2012.04.006>

593 Wittbrodt, B. T., Glover, A. G., Laureto, J., Anzalone, G. C., Oppliger, D., Irwin, J. L., & Pearce, J. M.
594 (2013). Life-cycle economic analysis of distributed manufacturing with open-source 3-D
595 printers. *Mechatronics*, 23(6), 713–726. <https://doi.org/10.1016/j.mechatronics.2013.06.002>

596 Yang, Z., Yang, H., & Yang, H. (2017). Effects of sucrose addition on the rheology and microstructure
597 of κ -carrageenan gel. *Food Hydrocolloids*, 75, 164–173.
598 <https://doi.org/10.1016/j.foodhyd.2017.08.032>

599 Zhu, F., Cheng, L., Yin, J., Wu, Z. L., Qian, J., Fu, J., & Zheng, Q. (2016). 3D Printing of Ultratough
600 Polyion Complex Hydrogels. *ACS Applied Materials and Interfaces*, 8(45), 31304–31310.
601 <https://doi.org/10.1021/acsami.6b09881>

602

603 **Figure captions**

604 **Figure 1.** Schematic view of 3D printing setup to promote the in situ temperature-induced
605 gelification.

606 **Figure 2.** Star-shaped 3D printed κ -carrageenan gel model, (a) before and (c) after introducing the air
607 distributor cooling ring (b).

608 **Figure 3.** Evolution of storage (G') and loss (G'') moduli with angular frequency of 3D printed and
609 conventionally prepared κ -carrageenan gel samples, as a function of (a) temperature, (b) layer
610 height and (c) printing speed.

611 **Figure 4.** Values of A and z parameters as a function of printing variables at different levels of (a,b)
612 layer height, (c,d) printing speed and (e,f) hotend temperature.

613 **Figure 5.** Time-temperature relationship for obtaining, by means of 3D printing, κ -carrageenan gels
614 with an imposed A value of $12000 \text{ Pa}\cdot\text{s}^{1/z}$.

615

616 **Table 1.** Independent variables studied and their normalised values used in the experimental design.

617

Printing speed [mm/s]	Hotend temperature [°C]	Layer height [mm]	Normalised value
10.0	50	0.06	-1.682
20.1	62	0.11	-1
35.0	80	0.18	0
49.9	98	0.25	1
60.0	110	0.30	1.682

618

619

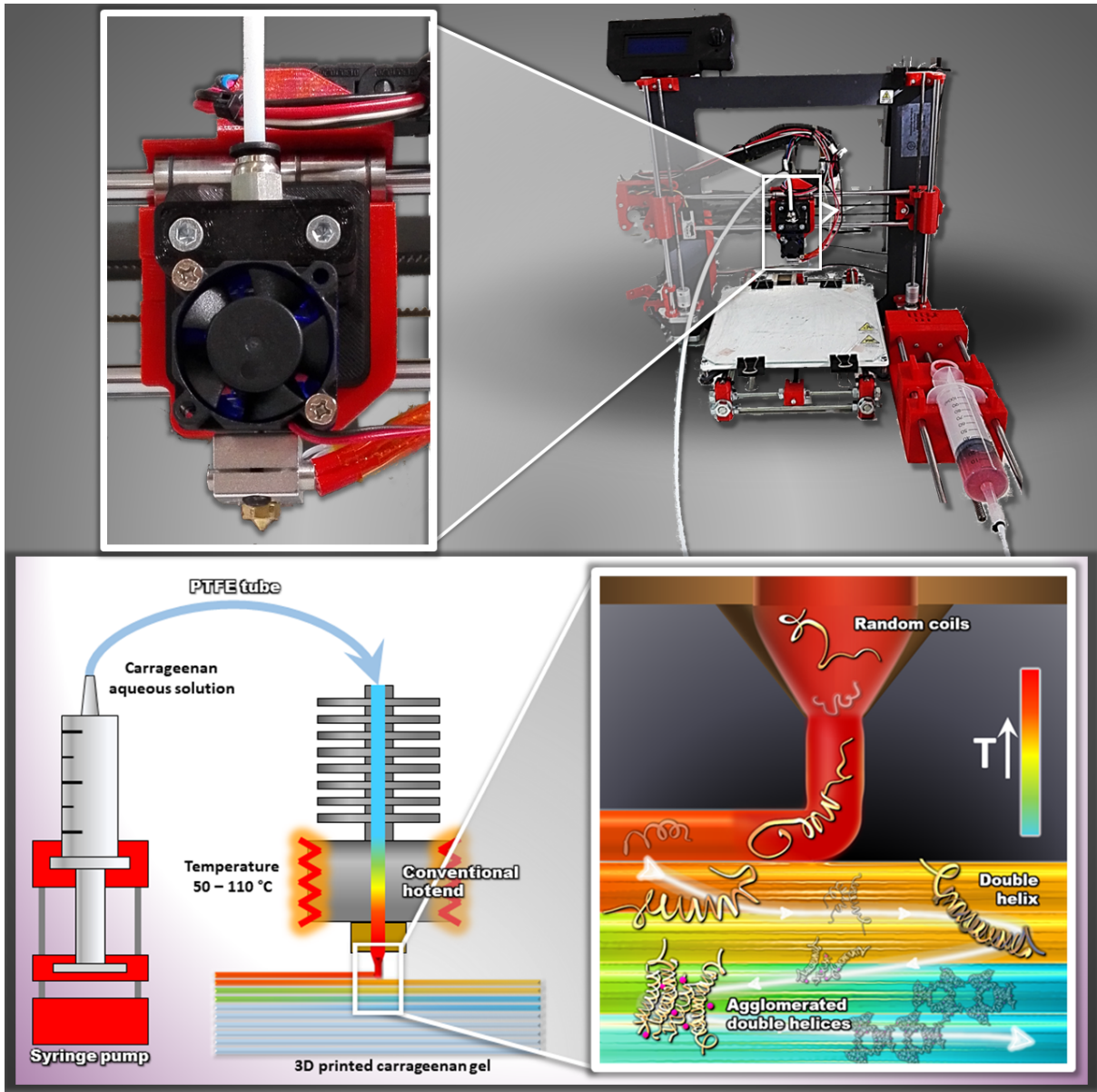
620 **Table 2.** Fitting parameters A and z obtained for each case study as a function of printing variables.

621

Case	Normalised values of independent variables			A [Pa·s ^{1/z}]	z	R ²
	S	T	H			
A	-1	-1	-1	13859.7	17.22	0.99
B	1	-1	-1	9719.1	17.89	0.99
C	-1	1	-1	18665.8	16.02	0.99
D	1	1	-1	12893.5	16.52	0.99
E	-1	-1	1	7444.0	13.91	0.97
F	1	-1	1	1662.3	12.11	0.95
G	-1	1	1	11754.5	15.36	0.98
H	1	1	1	7643.3	13.38	0.98
I	-1.682	0	0	15772.4	18.39	0.99
J	1.682	0	0	9593.0	15.44	0.99
K	0	-1.682	0	878.6	12.52	0.97
L	0	1.682	0	14506.7	13.28	0.99
M	0	0	-1.682	15125.7	18.19	0.99
N	0	0	1.682	7919.0	14.82	0.99
O1	0	0	0	10409.2	17.74	0.99
O2	0	0	0	9359.1	17.21	0.97
O3	0	0	0	10130.2	21.64	0.98
O4	0	0	0	11920.5	17.18	0.99
O5	0	0	0	11651.2	15.97	0.98
O6	0	0	0	11147.4	15.92	0.98

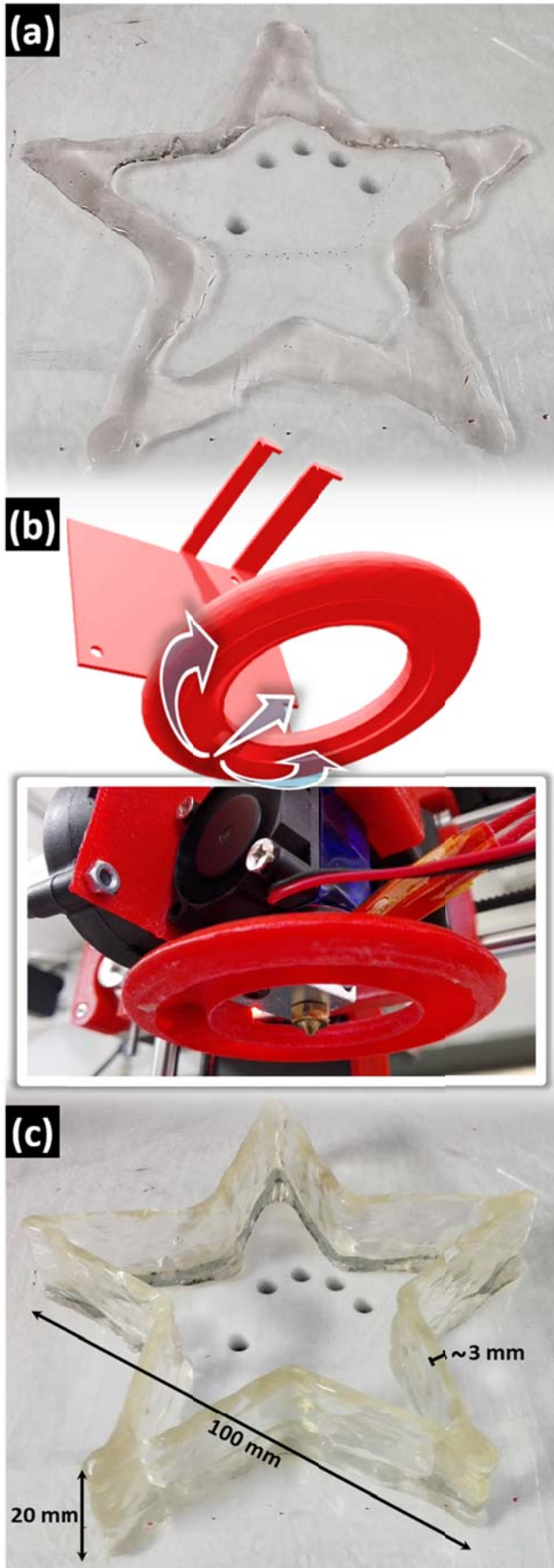
622

623



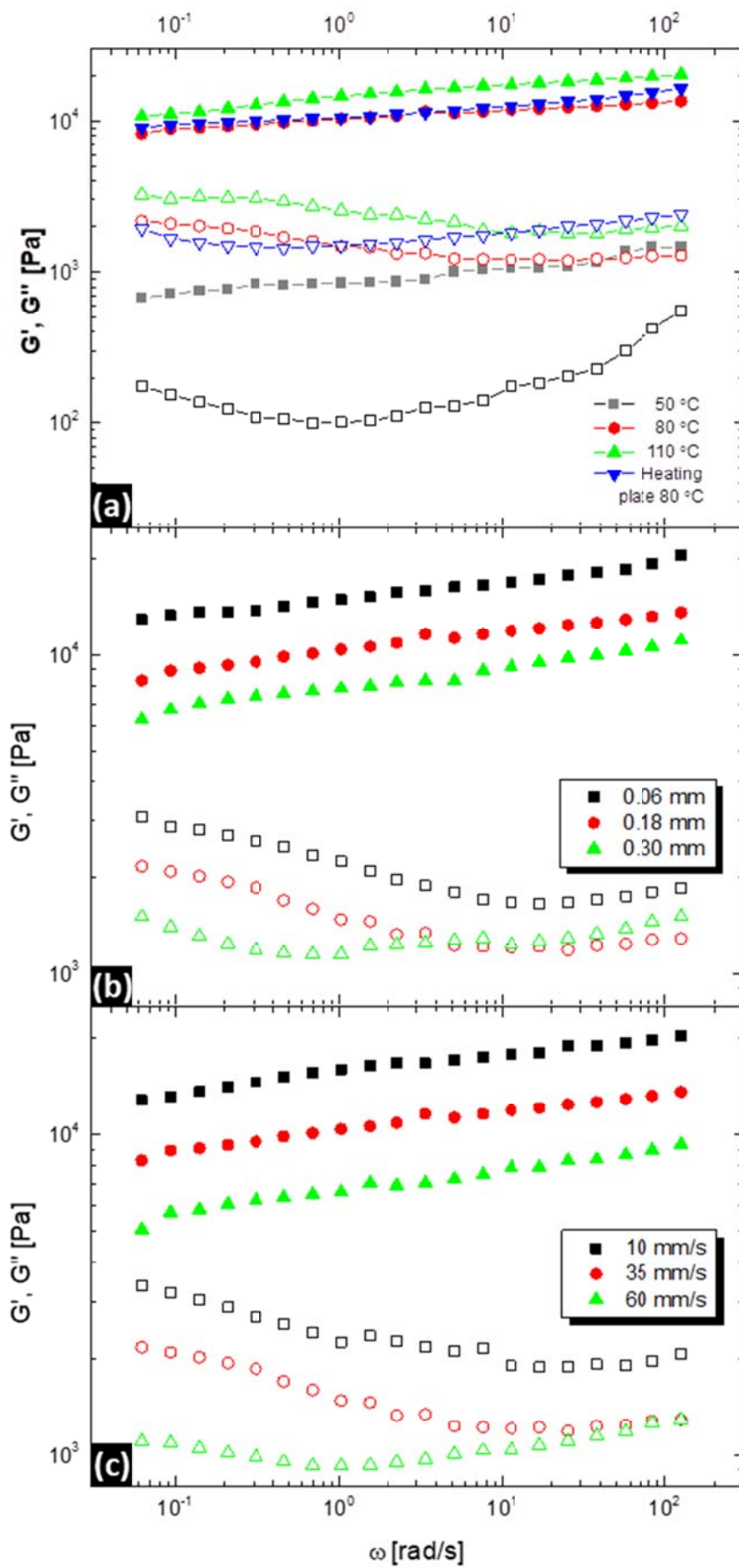
624

625 Figure 1.



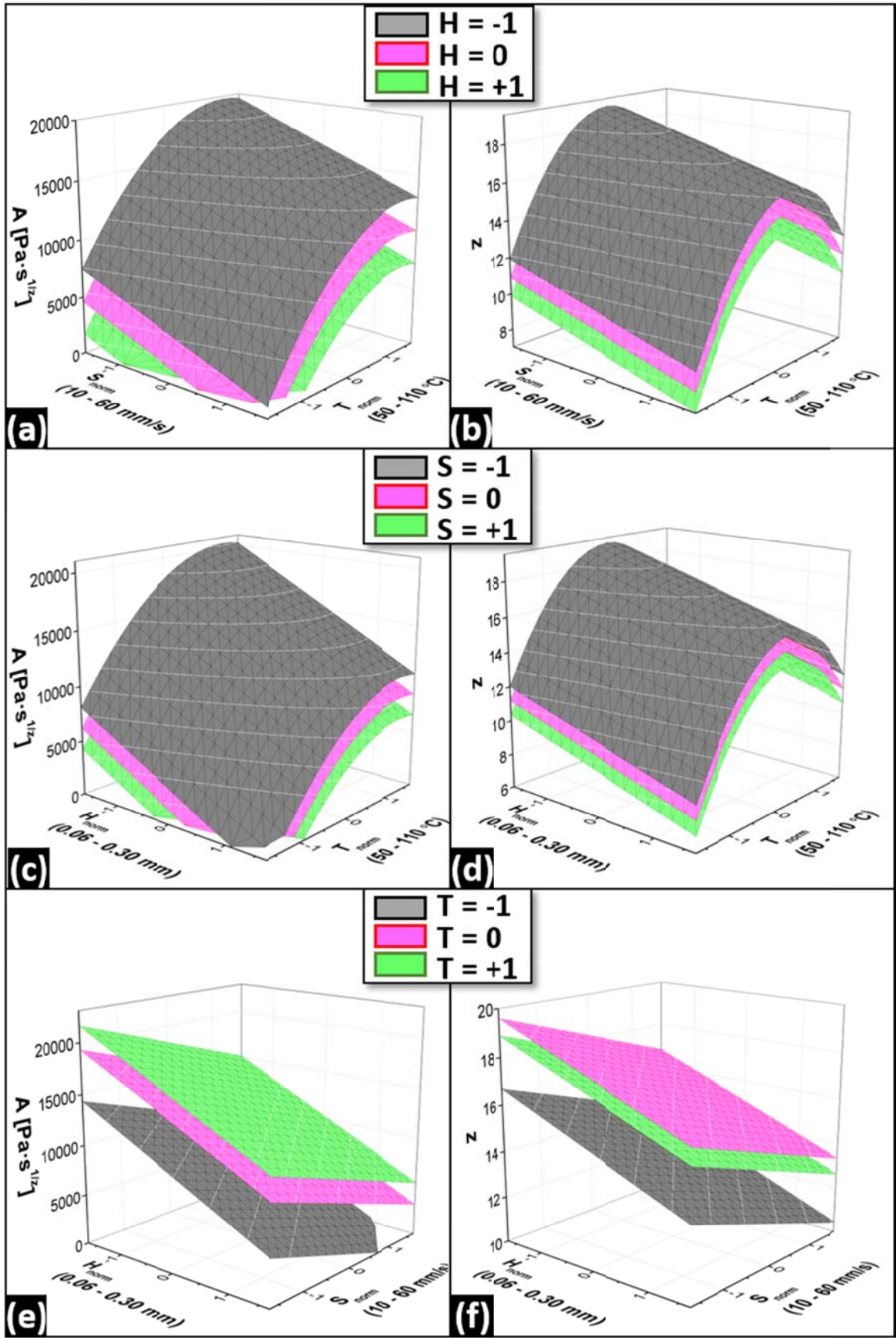
626

627 Figure 2.



628

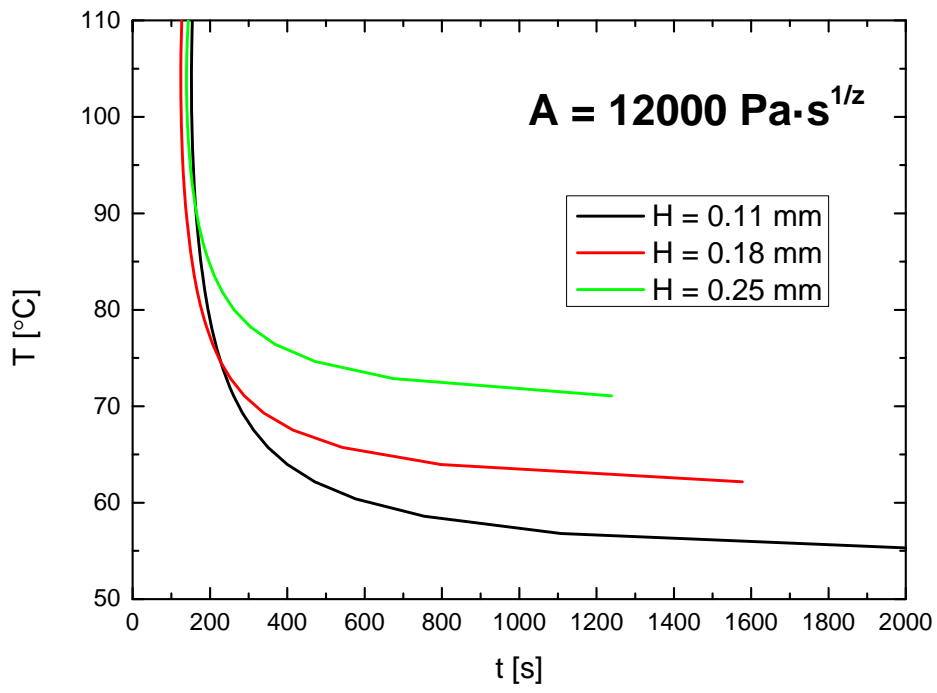
629 Figure 3.



630

631 Figure 4.

632



633

634 Figure 5.