

Original Article

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Microphytoplankton biometry as prospective vector for sulphide ore deposits: a case study in the Iberian Pyrite Belt

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Abstract

The Iberian Pyrite Belt contains one of the largest accumulations of massive sulphide deposits on Earth. Many of these deposits are hosted by latest Devonian black shales rich in terrestrial and marine palynomorphs. Among the marine fossils, the most abundantly reported species is *Maranhites mosesii*. By means of a multi-analytical methodology, including (1) biometry of specimens, (2) TOFSIMS imaging and spectral analysis of selected specimens and (3) host-rock geochemistry, we detected that cysts of *M. mosesii* are smaller and lighter in massive sulphide-generating environments than in coeval non-massive sulphide-generating environment. Cysts of *M. mosesii* sank after encystment and matured in the seafloor of different subbasins affected by disparate anoxic conditions. The specimens that matured in anoxic settings enriched in pollutants, like Arsenic (As) and Lead (Pb), were smaller and lighter than those from non-polluted anoxic environments. Their organic walls were also enriched in As. Neither the anoxia nor the pollutants prevented the proliferation of *M. mosesii*, as this was the most abundant phytoplanktonic species in all environments. To explain this, we suggest that this species developed a successful adaptive mechanism that might involve anaerobic metabolic interchange with the surrounding oxygen-depleted media and high levels of tolerance to stressors. Whatever the reason, it entails a causal relationship between cyst size and seafloor environmental conditions. In consequence, the biometry of *M. mosesii* can be envisaged as a promising vector for sulphide deposit exploration in the Iberian Pyrite Belt.

1. Introduction

The Iberian Pyrite Belt (IPB), located at the southwest margin of the Iberian Peninsula, contains one of the largest accumulations of massive sulphide deposits in the world. The original reserves of this Devonian-Carboniferous basin have been estimated at 1700 Mt of massive sulphides and 300 Mt of stockwork-type mineralization. Ores are primarily composed of pyrite with minor amounts of base metal sulphides (chalcopyrite, sphalerite and galena), other sulphides (including arsenopyrite) and sulfosalts. Besides Cu, Zn and Pb, important reserves of Sn, Au and Ag have been also documented. Mining exploitation in the region dates back over 5000 years. During this time, 82 sulphide deposits have been mined and several other indications have been documented (Pinedo Vara 1963; Sáez *et al.* 1996; 1997; 2024; Leistel *et al.* 1998; Tornos *et al.* 2000; Almodóvar *et al.* 2019; Sáez *et al.*, 2024).

At the latest Devonian, the environmental conditions of the hitherto stable IPB were drastically conditioned by the basin breakup and compartmentation associated with the onset of the Variscan cycle in the region (Moreno *et al.*, 1996; Soriano & Casas, 2002; Oliveira *et al.* 2004, 2019). The resulting IPB configuration consisted of a puzzling framework of coeval sub-basins filled with black shales, variable amounts of volcanic rocks and, on many occasions, large accumulations of massive sulphides (Moreno *et al.* 1996; Leistel *et al.* 1998; Sáez *et al.* 1999). These sub-basins, currently obscured by tectonic overprint, achieved disparate environmental conditions, and their black shales generally contain abundant spores and organic-walled microphytoplankton (González *et al.* 2005a; 2005b; Sáez *et al.* 2008; Pereira *et al.* 2021; 2023; Mendes *et al.* 2024). Among the marine organisms, the most prominently reported species is *Maranhites mosesii* (Sommer) Brito 1967 emend Gonzalez 2009.

Other microphytoplankton species found in the IPB, including acritarchs and prasinophyte microalgae, can be locally abundant, but for yet unknown reasons *M. mosesii* is by far the most widely and abundantly reported species in the region (González *et al.* 2005; González, 2009; Mendes *et al.* 2024).

According to González (2009), this microphytoplankton species exhibits high morphological variability linked to its ontogenetic development. However, while intra-sample size variations may be attributed to this growth trend, *M. mosesii* specimens from the IPB show significant inter-sample size disparities that are difficult to explain exclusively by this phenomenon. The present study provides the biometric analysis of populations of *M. mosesii* obtained from

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different localities, specifically from coeval massive sulphide-generating environments (MSGE) and non-massive sulphide-generating environments (non-MSGE). Geochemical analysis of the hosting black shales and Time-of-Flight Secondary Ion Mass Spectrometry (ToF-SIMS) spectra of a selection of specimens representative of each environment provide insights into the origin of this morphological disparity. The casual/causal relationship between morphological variation and presence/absence of massive sulphides in the hosting environments is thus explored.

To date, the information provided by the palynological analysis of the black shales hosting the IPB massive sulphides has been instrumental in biostratigraphically constraining the oldest mineralization event. Miospore assemblages invariably attributed to the uppermost Devonian Biozone of Western Europe, *Retispora lepidophyta-Verucosporites nitidus* (LN) by Streel et al. (1987), have been reported in the shales hosting the massive sulphides of Aznalcóllar (Pereira et al. 1996), Neves-Corvo (Pereira & Pacheco, 2004, 2021; Mendes et al. 2024), Tharsis (González et al. 2002), Sotiel Coronada (González et al. 2006), Torerera, Las Herrerías (Sáez et al., 2008), Caveira (Matos et al. 2014), Semblana-Rosa Magra and Monte Branco (Pereira et al. 2021). The multi-analytical approach here proposed represents a step forward, as it examines the viability of considering the morphology of *M. mosesii* as a prospecting method to discriminate between mineralized and barren sequences within the IPB.

2. Geological setting

The IPB is the largest and most representative domain comprising the South Portuguese Zone in the southern margin of the Iberian Variscan Massif (Figure 1a). Its stratigraphic record extends from Middle Devonian to Late Mississippian (González et al. 2003; Mendes et al. 2020; 2024; Moreno & González, 2004; Pereira et al. 2021) comprising three lithostratigraphic units (Schermerhorn, 1971). From footwall to hanging wall, these are: Phyllite-Quartzite Group (PQ Group), Volcanic-Sedimentary Complex (VSC) and Culm Group (referred to in Portugal as Baixo Alentejo Flysch Group) (Figures 1a and 2). The PQ Group consists of a thick detrital sequence of shales and sandstones, Givetian–latest Famennian in age, deposited in a stable and shallow siliciclastic platform, eventually affected by storms (Sáez & Moreno, 1997; Matos et al. 2014; Oliveira et al. 2013; 2019). To the top, the series incorporates isolated limestone bodies (see, for example, van den Boogaard, 1963; van den Boogaard & Schermerhorn, 1981; Albardeiro et al. 2020; Mendes et al. 2020) and conglomerates, as well as mega debris-flow and fan-delta deposits, all in relation with the instability affecting the IPB during early propagation of the Variscan cycle (Moreno et al. 1996). Such instability resulted in the Late Devonian fragmentation and compartmentation of the IPB basin. The transition between the PQ Group and the VSC is commonly associated with a black shale sequence rich in organic matter including minor interbedded volcanics. This so-called anoxic sequence (Moreno & González, 2004) hosts many of the large massive sulphide deposits from the southern part of the region, including Tharsis, Sotiel Coronada-Migollas, Aznalcóllar-Los Frailes, Neves-Corvo, Lousal and Carreira. Above the anoxic sequence, the VSC incorporates a diversity of Mississippian sedimentary volcanic and subvolcanic rocks, mafic and felsic in nature, displaying frequent thickness variations and abrupt lateral and vertical facies changes. This unit is interpreted in relation to the transtensional effect of the early syn-orogenic stage occurring

in a heterogeneous and compartmentalized basin (Moreno et al. 1996; Martín-Izard et al. 2017; Oliveira et al. 2019).

Sulphide deposits from the north and intermediate zones of the IPB are mostly associated with volcanic and volcanoclastic rocks of the VSC. The geochronological constrain of these deposits is still poor, but limited available data indicate a late Tournaisian age (Barrie et al. 2002; Dunning et al. 2002; Albardeiro et al. 2023). Considering all the biostratigraphic and geochronological age data, the existence of two mineralizing pulses (late Famennian and late Tournaisian) or a single continuous metallogenic episode extending from late Famennian to late Tournaisian is still under debate (Albardeiro et al. 2020; 2023; Sáez et al. 2024).

The contact between the VSC and the overlying Culm Group is marked by another shaly sequence, the Basal Shaly Formation (Moreno & Sequeiros, 1989), referred to in Portugal as the Brancanes Formation (Oliveira et al. 2013, 2019). This unit comprises reworked material from the uppermost VSC intervals, along with siliciclastic deposits linked to the initial post-volcanic turbiditic pulses in the region. Above this formation, the Culm Group comprises the syn-orogenic infill of the IPB basin (Moreno, 1993; Moreno et al. 1996; Oliveira, 1990; 2019). During the Asturian phase of the Variscan orogeny, the entire succession was deformed following a thin-skinned tectonic style (Silva et al. 1990).

The four sampling sites selected in this study are stratigraphically located within the anoxic sequence transitional between the PQ Group and VSC (Figure 2). All samples collected in these sites were palynostratigraphically analyzed and attributed to the uppermost Famennian LN Miospore Biozone (Streel et al. 1987) of Western Europe. Two of the studied sites, Tharsis and Sotiel, are located in MSGE, whereas the other two, Águila Range and Calañas, are from non-MSGE (Figure 1). Samples from non-MSGE are outcrop samples. Those from Tharsis and Sotiel are, respectively, from an open pit and from an underground mine. The stratigraphic location of the samples studied in each locality is illustrated in Figure 1b. Details of the biostratigraphic analysis carried out on each site can be seen in González et al. (2002, 2005, González et al., 2006) and Moreno et al. (2003).

3. Materials and methods

The analysis performed on all the samples includes biometry, Time-of-Flight Secondary Ion Mass Spectrometry (TOF-SIMS) of selected *M. mosesii* specimens and geochemical analysis of the hosting black shales. Biometry and TOF-SIMS were preceded by a palynological extraction procedure, which essentially followed the standard tri-acid digestion method described by Wood et al. (1996). This comprises initial disaggregation of approx. 35 g of sample, immersion in HCl (36%) to remove carbonates, dissolution of silicate minerals with hot HF (40%) and a second immersion into hot HCl (36%) to eliminate fluorides and remaining carbonates. After neutralization, the organic residue was oxidized with Schulze solution for variable time intervals. The oxidized residue was mounted in microscope slides using Cellosize as dispersant and Elvacite as mounting medium. Preservation status of the selected samples was optimal for palynological identification. After biostratigraphic recognition and palynofacies analysis, based on 350 counts, the biometry of *M. mosesii* was performed selecting 120 specimens per sample, when possible. The morphological features measured were: vesicle diameter, equatorial pads, equatorial embayment, lateral thickenings and translucent bladders. Details on these structural elements can be seen in Figure 3 and in González (2009). Biostratigraphy, palynofacies and

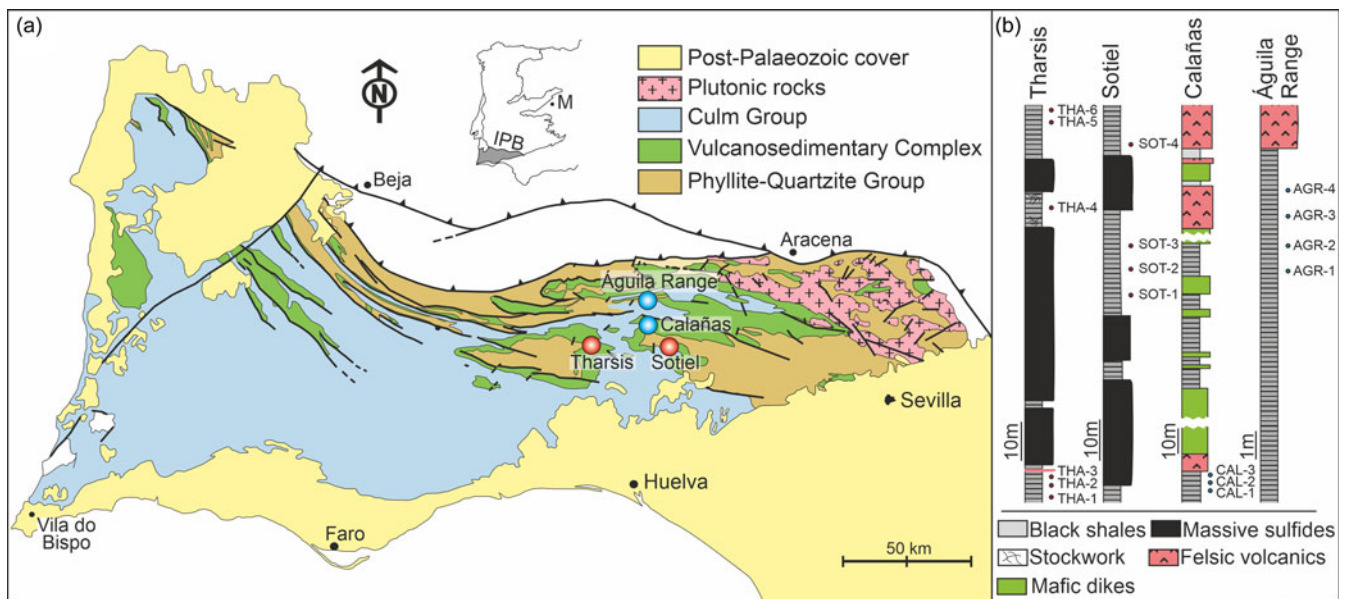


Figure 1. (a) Geological sketch map of the Iberian Pyrite Belt (IPB) with indication of the four sub-environments/sub-basins considered in this study. M, Madrid; IPB, Iberian Pyrite Belt. Red circles indicate MSGE. Blue circles indicate non-MSGE. (b) Stratigraphic logs at these localities showing the location of the studied samples. All the samples are latest Famennian (LN Biozone) in age.

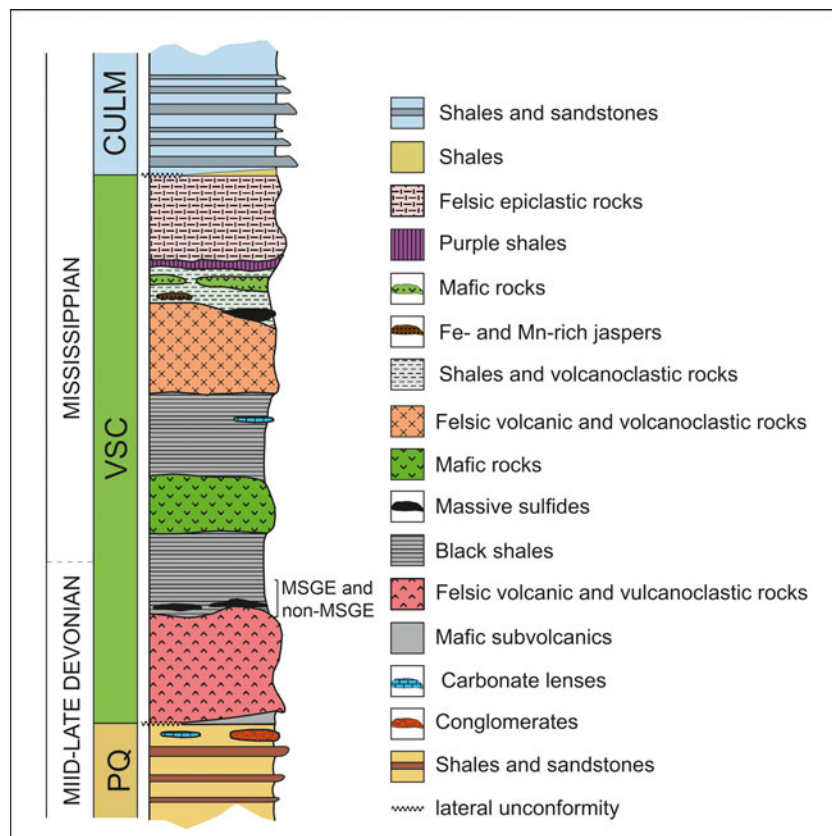


Figure 2. Synthetic stratigraphic log of the IPB.

biometric analyses were performed in the Department of Earth Sciences, University of Huelva, using a Nikon Labaphot-2 microscope equipped with the digital camera Nikon DS-Fi1 (Software NIS Elements, F Package, 3.22.00). Processed and

remaining unprocessed samples are deposited in the collections of the Earth Sciences Department, University of Huelva.

For the TOF-SIMS analysis, 20–40 specimens per sample of *M. mosesii* were picked with a micromanipulator needle and

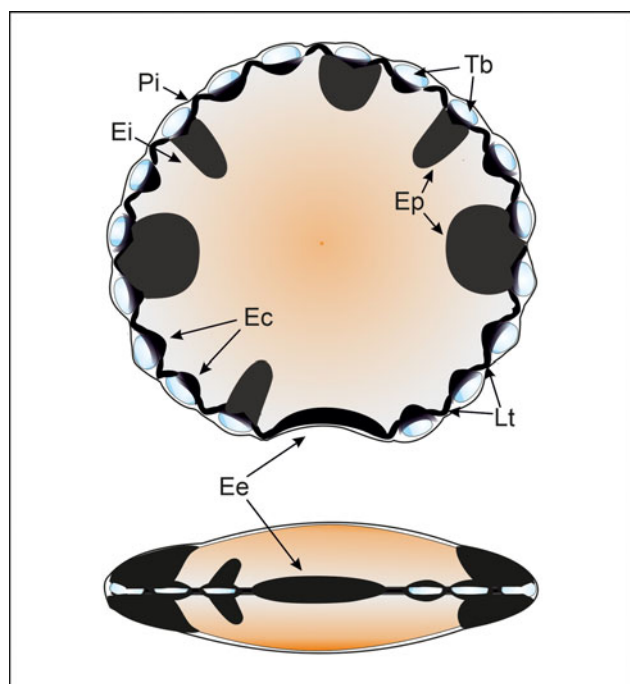


Figure 3. Polar and equatorial scheme views of *M. mosesii* after González (2009) with indication of its main equatorial structures: (Pi) Perieilyma. Outer vesicle wall; (Ei) Endeilyma. Inner vesicle wall; (Ep) Equatorial pads. Solid, protrusive discrete to rarely interconnected structures, circular to subcircular in outline; (Ec) Embracing cells. Dark equatorial structures, elliptical to subcircular in outline, that enclose translucent bladders and are connected by lateral thickening; (Lt) Lateral thickening. Small, solid, circular, subcircular or arcuate structures that connect embracing cells; (Tb) Translucent bladders. Hollow, rounded to elliptical equatorial structures, enclosed by embracing cells, that progressively enlarge during vesicle maturation; (Ee) Equatorial embayment, normally thickened, that may or may not be accompanied by embracing cells or equatorial pads.

mounted in a double-sided Cu tape. Measurement and chemical imagery were then performed using a TOF-SIMS IV instrument (ION-TOF GmbH, Germany) located at the Scientific and Technological Research Assistance Centre of the University of Vigo, Spain. The analysis was carried out with a primary ion beam of 25 KeV Bi³⁺ (pulsed current between 0.12 and 0.15 pA) scanning over a 500 × 500 μm² area for 100–150 s. The Primary Ion Dose Density was settled to be below the static limit (1013 ions/cm²). A pulsed electron flood gun source was employed for surface build up charge compensation.

Spectra (m/z 0–1000) and chemical maps of selected m/z values (scans acquisition repeated 75 times) were collected for all samples in positive and negative polarity. Positive and negative ion spectra were internally mass-calibrated using CH₃⁺, C₂H₃⁺, C₃H₅⁺ and C₅H₁₄NO peaks for positive ions, and CH[−], C₂H[−], C₄H[−] and C₁₈H₃₅O₂[−] peaks for negative ions, respectively. Deviation between theoretical and measured masses was always below 100 ppm. Combining the elements described in the literature as potential contaminant for phytoplankton with those that could be mobilized in oxygen-depleted and/or hydrothermally influenced environments, the list of major and trace elements searched in TOF-SIMS profiles includes Mg, Al, Si, Cr, Fe, Co, Zn, As, Cd, Cs, Ba, Ce, Cu, Hg and Pb.

Chemical composition of shales was obtained after milling on agate mortar and split into 20 g fractions at the University of Huelva laboratories. Geochemistry was characterized by analyzing major and trace elements together with carbon and sulphur in a

fully certified commercial laboratory (ACME, Analytical Laboratories, Vancouver, Canada). Total abundances of major oxides were determined by ICP emission spectrometry. Rare Earth and refractory elements were analyzed by ICP mass spectrometry. Both procedures followed Li-metaborate/tetraborate fusion and dilute nitric digestion of samples weighing 0.2 g. Sample splits of 0.5 g were analyzed for a selection of metals and semi-metals including Cu, Pb, Zn, As, Se and Hg after aqua regia digestion. Sulphur, total C and organic C were determined by Leco analysis.

4. Results

4.a. *Maranhites mosesii*

The microphytoplankton species *M. mosesii* extends from Early Devonian to Mississippian. It is widely distributed throughout Gondwana and southern margin of Laurussia and is generally associated with shallow marine depositional environments, relatively close to emergent landmasses (González, 2009). Morphologically, it consists of a vesicle, originally discoidal, circular to subcircular in outline, 40 to 200 μm in diameter and two-layered. The inner layer (endeilyma), noticeably thicker than the outer layer (perieilyma), contains well-differentiated structural elements at equatorial positions, including solid pads, embracing cells, transparent bladders, lateral thickenings and an equatorial embayment (Figure 3).

The ontogenetic trend detected in *M. mosesii* (González, 2009) entails increasing vesicle size and continuous transformation of these equatorial structures. Growth trends also involving vesicle enlargement have been described in cyst-forming prasinophyte species during their non-motile stage (Tappan, 1980; Knoll *et al.* 1991; Teyssède, 2006; Taylor *et al.* 2009). As in prasinophytes, the progressive cyst enlargement of *M. mosesii* cannot be solely attributed to the consumption of internal reserves, necessitating metabolic exchange with the surrounding environment.

The biometric analysis of *M. mosesii* shows that the average equatorial diameter and the number of pads-bearing specimens clearly differ between MSGE and non-MSGE (Figure 4). In samples from the two MSGE selected in this study (Figures 1 and 2), the average equatorial diameter of *M. mosesii* specimens ranges from 68.2 to 73.6 μm and 70.7 to 76.7 μm, respectively, whereas in non-MSGE, this increases up to 81.7 to 92 μm and 86 to 102 μm. The results show that the specimens in MSGE are about 15% smaller than in non-MSGE (Figures 4 and 5). Considering the number of pad-bearing specimens, the percentage observed in MSGE ranges from 3.0% to 32.9%, whereas in non-MSGE, it varies between 43.2% and 52.8%. This indicates a reduced occurrence of pad-bearing specimens in MSGE compared to non-MSGE. There is only one sample from MSGE that, with 47.1% of pads-bearing specimens and 88.1 μm of average equatorial diameter, clusters better with those from non-MSGE. The rest of structural elements measured in *M. mosesii* evidence variations among samples with no clear tendencies or signs of clustering (Supplementary Table 1).

4.b. Inorganic element fixation during cyst stage

The elements assimilated by *M. mosesii* detected with TOF-SIMS spectra are shown in Figure 6. The most striking feature concerns the behaviour of As. This element is commonly detected, with variable intensity, in all but three MSGE samples, but it is absent or shows very low concentration in most of the samples from non-MSGE. Other elements, including Mg, Al, Si, Fe, Zn, Co, Cs, Cr, Ba and Pb, were detected virtually in all TOF-SIMS spectra, but with

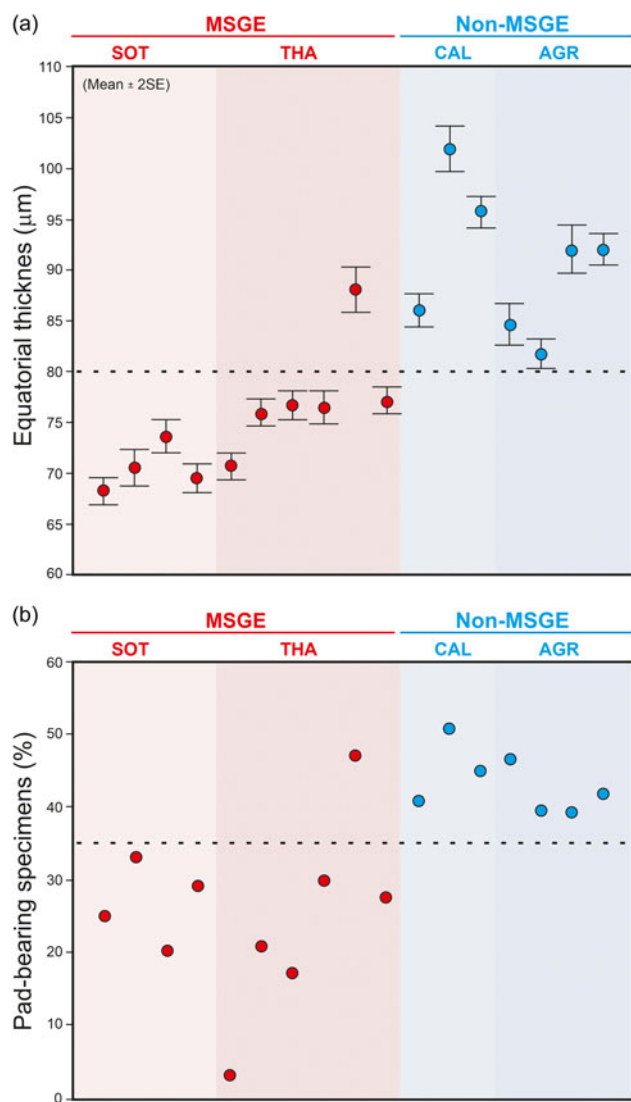


Figure 4. Average equatorial size (a) and percentage of pad-bearing specimens of *M. mosesii* (b) in samples from MSGE and non-MSGE. SOT, Sotiel; THA, Tharsis; CAL, Calañás; AGR, Águila Range.

variable intensities and no differentiation between MSGE and non-MSGE. Of these, Si shows intense peaks in all cases, Co is the element with the highest variability and Ba and Ce depict very similar behaviour, drawing a distribution pattern apparently not related to the environmental differentiation followed in this study. The rest of the elements analyzed, i.e., Cu, Hg, Ce and Cd, were absent, detected randomly or with very low intensity. The elemental chemical TOF-SIMS images (Figure 7) show that the level of enrichment in As, Fe and Al occasionally differs among specimens from a given sample, indicating that these elements were gradually assimilated by *M. mosesii* during cyst growth. Such contrasting occurrences exclude the influence of diagenetically controlled enrichment mechanisms. Other elements, including Si, Mg, Co, Cs and Pb, remain fairly constant in all the specimens of a given sample, suggesting that the enrichment mechanism was in these cases related to a generalized post-depositional process. At the level of specimen, the TOF-SIMS shows no compositional variations across the surface, indicating that the outer wall of *M. mosesii* has a uniform chemical structure and that the elemental

enrichment, either at cyst stage or during post-depositional assimilation, was also uniform.

4.3. Seafloor environment reconstruction

Geochemical analysis of the black shales hosting the cysts of *M. mosesii* indicates oxygen-deficient conditions according to Hatch and Leventhal (1992) and Hoffman *et al.* (1998), with the chemocline very close to the sediment-water interface (Jones & Manning, 1994) and no significant differences between MSGE and non-MSGE (Figure 8a and Supplementary Table 2). The alternative use of the U and Mo enrichment factors (Algeo & Tribovillard, 2009; Algeo & Li, 2020) yields similar results (Figure 8b). The absence of covariation between both factors and the consistently low absolute values of Mo_{EF} and U_{EF} for all samples, except for one ($U_{EF} = 7.32$), support the interpretation of generally oxic marine conditions, with the redox chemocline positioned near the water-sediment interface (Algeo & Tribovillard, 2009). For full discussion on redox condition of IPB sulphide-hosting environments see Sáez *et al.* (2011). Although the precise extension of the oxygen-depleted water column is not provided by these geochemical data, the abundance of organic-walled microphytoplankton, including *M. mosesii*, in all samples suggests a vertical zonation pattern with oxygenated water at least in the upper photic zone. Both acritarch cysts and prasinophyte phycocysts, the two microphytoplankton groups reported in the studied samples, are generated by the encystment of motile aerobic organisms that, for comparison with modern analogues, require a photic and oxygenated habitat for living (Wong *et al.* 2023).

The comparison of spider-like diagrams for black shales from the two environment sceneries considered in this study shows strong positive anomaly for As and Se in MSGE. Minor anomalies for P, Cu, Zn and Pb are also noticeable (Figure 8c). Analytical values for Cu, Pb and As normalized to TiO_2 are presented in Figure 8d. It shows that all but two samples from non-MSGE form a cluster near the Cu corner, whereas shales from MSGE are dispersed and relatively enriched in As and Pb.

In the IPB, the generation of sulfide deposits was associated with highly efficient hydrothermal systems (Sáez *et al.* 2024). High-temperature hydrothermal mineralizing fluids co-transported reduced sulphur and metals from deeper to near-surface zones. Under this scenario, the occurrence of Cu, As and Pb in MSGE seafloor, along with the other metals that ultimately contributed to the formation of the massive sulphide deposits, would be expected.

In non-polluted marine environments, Cu is generally used as proxy for bioproductivity (Tribovillard *et al.* 2004; Riquier *et al.* 2006), while As and Pb are considered highly poisonous for living organisms (Davies, 1979; Sharma & Sohn, 2009).

5. Discussion

The conjugate analysis of the three techniques employed provides information about the interaction of *M. mosesii* with their hosting environment. Although the morphological variability detected here is linked with the growth trend previously accounted for this species (González, 2009), specimens reported in MSGE are systematically smaller and lighter than those from non-MSGE. Besides, their external wall is enriched in As, a poisoned element more abundant in MSGE than in non-MSGE MSGE that is common in IPB massive sulphide and stockwork mineralization in the form of arsenopyrite and tennantite (Almodóvar *et al.* 2019).

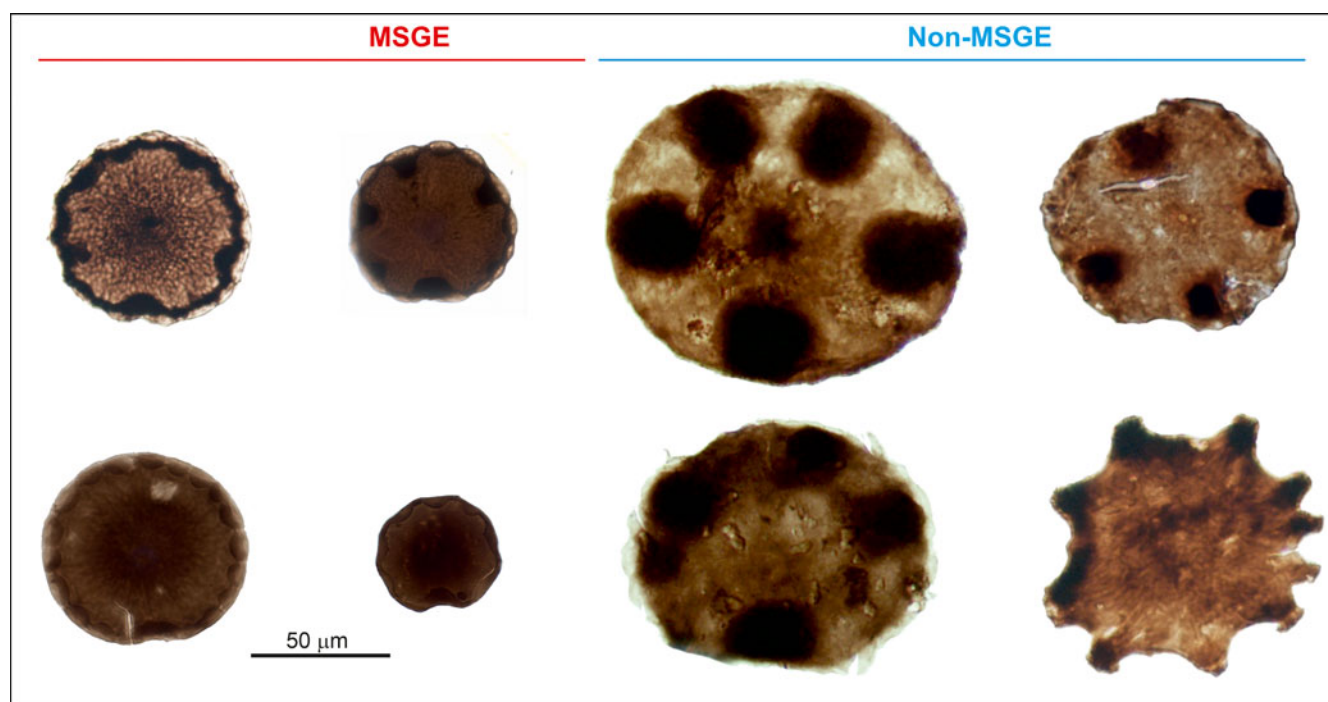


Figure 5. Specimens of *M. mosesii* recovered in MSGE and non-MSGE showing clear differences in equatorial size.

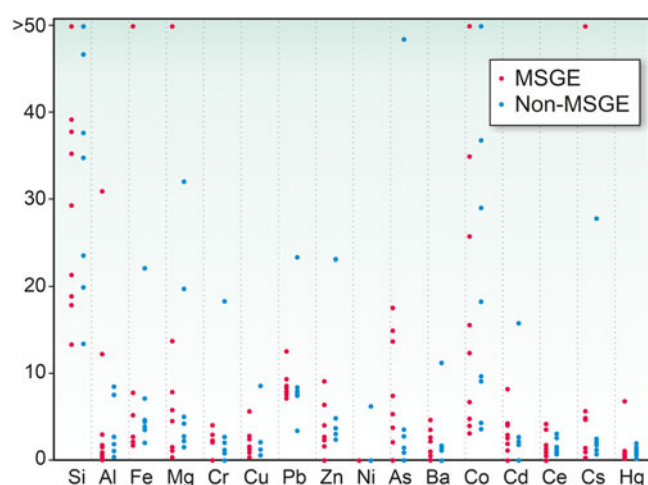


Figure 6. Elements detected in *M. mosesii* from MSGE and non-MSGE based on TOF-SIMS spectra. Vertical scale represents enrichment factor with respect to the holder (adhesive tape). Spectra normalized to PDMS peaks (m/z 147 and 207) identified in the holder tape.

This indicates that cysts of *M. mosesii* sank after encystment, maturing at the bottom of the different sub-basins. There, the disparate environmental conditions conditioned the cyst maturation.

In living, cyst-producing dinoflagellates, the intraspecific morphological variability has been largely associated with changes in temperature, salinity, nutrient levels or pollutants (Dale, 1996; Ellegaard *et al.* 2002, 2017; Pospelova *et al.* 2002, 2005). In the fossil record, morphological perturbations of dinocysts and acritarchs have been also associated with local changes in salinity (Dybkjær, 2004), but more commonly with global factors such as fluctuations in the C cycle (Delabroye *et al.* 2012; Munnecke *et al.* 2012) or

expansion of oceanic anoxic events (Vandenbroucke *et al.* 2015). Nevertheless, the variability detected in acritarchs and dinoflagellates, either living or fossil, is understood as the interaction of their parent flagellate cells with the surrounding environment (Kremp *et al.* 2009; Lundholm *et al.* 2011). During the non-flagellate phase, the metabolic activity drastically decreases, remaining, in most cases, at the expense of internal carbohydrate reserves (Binder & Anderson, 1990). Thus, the resting cysts of acritarchs and dinoflagellates virtually operate as closed systems, sensitive to variations in temperature, light and redox conditions (Anderson *et al.* 2005; Brosnahan *et al.* 2020), but not interacting or interacting minimally with their hosting environment.

In the case of living or fossil prasinophytes, studies about intraspecific morphological variations and differential growth rates are very scarce. The available data suggest that, although the morphology of the prasinophytes cyst, called phycmata, can be governed by global climatic factors (Spiridonov *et al.* 2017), it must be also influenced by the environmental conditions prevailing at maturation sites (Boalch & Mommaerts, 1969). There, the substantial phycmata enlargement (Wiebe *et al.* 1974; Parke *et al.* 1978; Colbath, 1983; Knoll *et al.* 1991; Tyson, 1995), which in current prasinophytes can represent an increase of up to 5000 times the size of the precursor flagellate cells (Tappan, 1980), requires of an extracellular energetic source (Pazio, 2016).

On the other hand, the effects of As on modern phytoplankton have been studied in detail. Although with different efficiency, these organisms are capable of bio-uptaking inorganic As, generally as arsenate (III), and subsequently reduce it to arsenite (IV) in order to form more complex and less toxic methylated organic compounds that are then excreted as a detoxification mechanism (Neff, 1997; Yamaoka *et al.* 1999; Goessler *et al.* 1997; Caumette *et al.* 2011; Giovannoni *et al.* 2019). They are even capable of assimilating small proportions of inorganic As. However, when they are exposed to anomalously high As

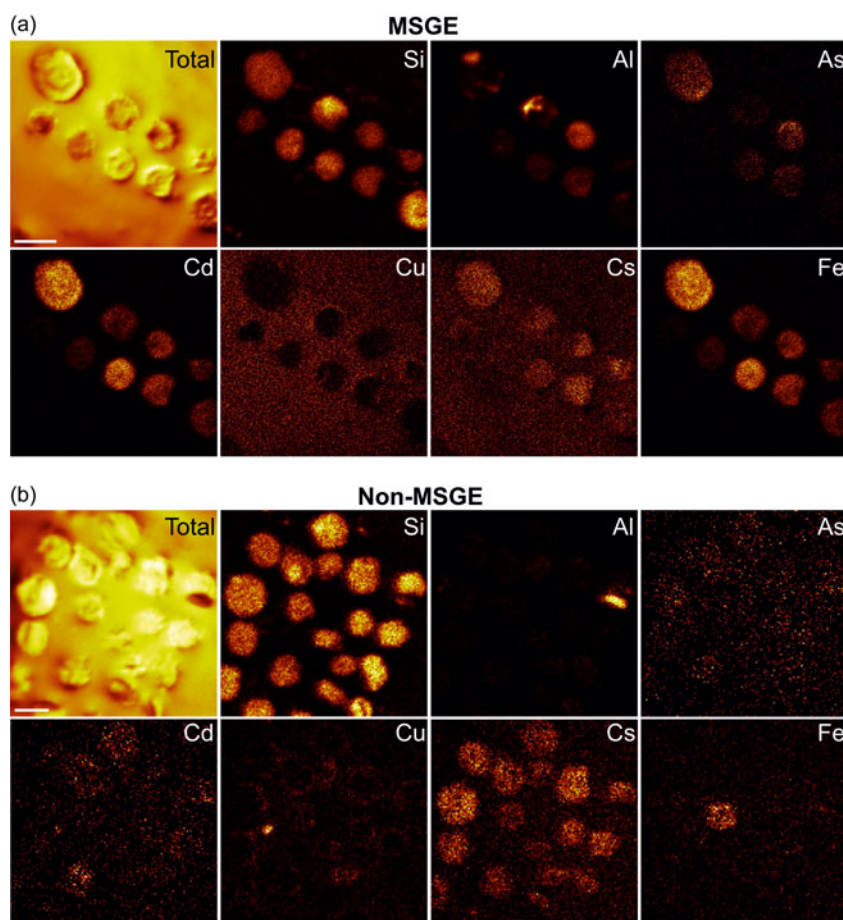


Figure 7. TOF-SIMS chemical images of selected elements detected in specimens of *M. mosesii* from sample THA-4, as representative of MSGE (a) and sample AGR_1, as representative of non-MSGE (b). Scale bars 100 μm.

concentrations, their maturation capacity can be affected and even their lethality increases (Planas & Healey 1978). This behaviour regarding As refers exclusively to non-cyst-forming species or to the flagellate phase of cyst-forming phytoplanktonic organisms. The drastic metabolic reduction in their cyst phase minimizes their element assimilation and their potential infection. This explains the absence of information in the available literature on the effect that As, or any other element, can produce on dinoflagellate and acritarch resting cysts. There are also no studies to our knowledge on the effect of the environment on current prasinophyte phycomata, despite in this case, its huge enlargement suggests metabolic activity beyond the consumption of internal carbohydrate reserves.

In the absence of comparative studies and considering the environmentally induced behaviour of *M. mosesii* here detected, we hypothesize that this species established an anaerobic interchange with the surrounding oxygen-depleted environment during cyst maturation and that As fixation was a by-product of such metabolic process. During maturation, the metabolic activity responsible for As assimilation gradually gained importance. This explains the different As concentrations detected among specimens of a given sample (Figure 7). In non-MSGE, the cysts of *M. mosesii* also contain As, although in minor proportion. This is explained by the ubiquitous occurrence of As in the marine realm, particularly in oxygen-depleted environments (Leventhal, 1991; Bodin *et al.* 2006; Liu & Cai, 2010). When cysts reached an optimum level of maturation, regardless of the level of As accumulated, they moved up in the

water column favoured by near-bottom and/or upwelling currents, and finally dispersed their offspring in well-oxygenated near-surface waters. Supporting this hypothesis is the fact that cyst-forming phytoplankton requires oxygenated environments for excystment and germination (Anderson *et al.* 1987; Kremp & Anderson, 2000; Ellegaard *et al.* 2017), and that specimens of *M. mosesii* having excystment structures, i.e., those that resuspended to the photic zone and settled on the bottom again, are in much smaller proportion than non-excysted cysts. The equatorial solid pads in this species (Figures 3 and 5) could have been used as energy storage devices or as structures within a floating/sinking system. Their consumption or detachment, joined with the development and progressive enlargement of the equatorial, hollow, transparent bladders that characterize the latest growth stages in this species (González, 2009), favoured the final resuspension of mature cysts. The elevated concentration of As in MSGE decreased the metabolic interchange with the surrounding medium or accelerated the cysts maturation, and with it their resuspension, in order to attenuate, or diminish, the exposure to a hostile environment. The reduced equatorial diameter and small amount of pad-bearing cysts were the consequences of this process.

The common to elevated occurrence of *M. mosesii* cysts in most MSGE samples suggests that such adaptive mechanism was effective. However, the quantitative analysis of the relative proportion of acritarchs, prasinophyte phycomata and *M. mosesii* specimens (Figure 9) shows that MSGE samples are mostly dominated by acritarchs whereas non-MSGE samples are mostly

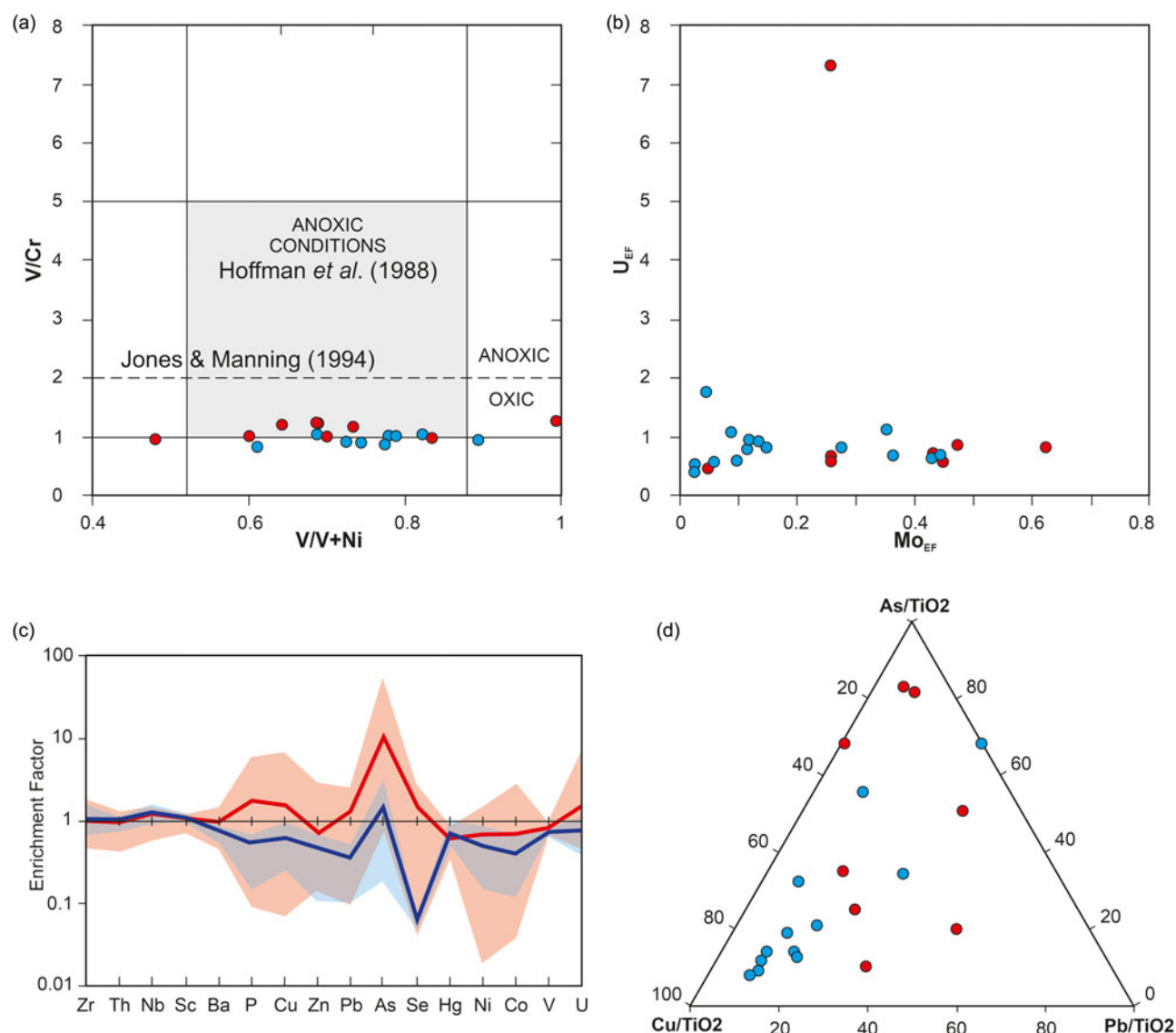


Figure 8. (a) Bivariate plot of the black shale samples from MSGE (red circles) and non-MSGE (blue circles) in the diagram V/Cr vs V/(V+Ni). Limits of environmental conditions after the Hoffman *et al.* (1998) and Jones and Manning (1994); (b) Diagram U_{EF} vs Mo_{EF}; (c) PAAS-normalized spider-like diagrams for maximum, minimum and average values (bold lines) of black shale samples. Red and blue lines represent samples from MSGE and non-MSGE, respectively; (d) Ternary diagram showing the relative proportion of Cu-As-Pb normalized to TiO₂. Red and blue circles for MSGE and non-MSGE, respectively.

dominated by *M. mosesii*. In addition, the percentage of this species is most fluctuating in MSGE than in non-MSGE samples. This means that, although *M. mosesii* can be adapted to pollutant conditions, they germinate more abundantly in non-stressed environments.

Regardless of whether cyst size reduction in *M. mosesii* is an adaptive mechanism or a limitation resulting from growing under hostile conditions, a causal relationship between cyst size and hosting environment is clearly established. This means that, in the IPB, the biometric analysis of *M. mosesii* specimens provides an effective tool for distinguishing late Devonian black shales sequences associated with massive sulphides from barren sequences (without massive sulphides). To date, the contribution of palynology to the study of the IPB basin has primarily focused on the detailed biostratigraphic analysis of shale sequences belonging to its three chronostratigraphic units, with special emphasis on the VSC sequences hosting the massive sulphides and

the coeval underlying PQ sequences cut by stockwork type mineralization (i.e., Pereira *et al.* 1996, 2004, 2021, 2023; González *et al.* 2002, González *et al.*, 2006; Mendes *et al.* 2020, 2024; Sáez *et al.* 2007). The present study goes a step further by aiding in the identification of potential targets for sulphide exploration in the region, paying special attention to the massive sulphide and stockwork-type mineralization hosted by black shales.

Finally, in terms of suprageneric affiliation, *M. mosesii*, and for extension the genus *Maranhites* (Brito 1965) emend González 2009, cannot be unquestionably assigned to the Prasinophyceae Class due to the absence of morphologically comparable extant counterparts. However, its preference for oxygen-depleted environments and its active metabolism during the cyst phase, unreported in living and fossils acritarch and dinoflagellates, reinforces the hypothesis by Tappan (1980), Colbath and Grenfell (1995) and Lé Hérisse *et al.* (2009) by which *Maranhites* may have prasinophyte affinity.

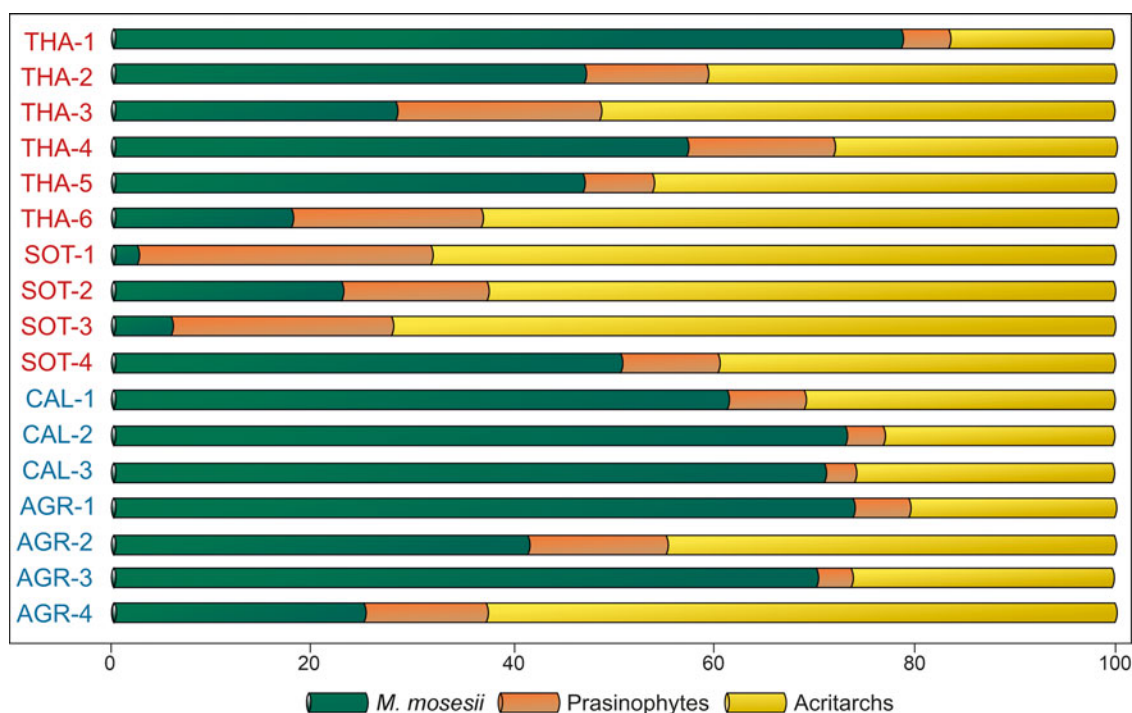


Figure 9. Quantitative analysis of the phytoplankton recovered in the studied samples (in percentages). Samples in red from MSGE. Samples in blue from non-MSGE. Location of samples is indicated in Figure 1.

6. Conclusions

The latest Devonian palynological assemblages from the IPB are unusually enriched in the microphytoplankton species *M. mosesii*. This species is abundant in both MSGE and non-MSGE environments. The geochemistry of the shales recovered in MSGE and non-MSGE environments indicates that both habitats are characterized by oxygen-depleted seafloor conditions and that most of the MSGE samples were enriched in pollutants, including As and Pb. The specimens of *M. mosesii* reported in MSGE were smaller and lighter than those from non-MSGE. Besides, the MSGE specimens depicted anomalous high values of As on their outer walls. This indicates that cysts of this species sank after encystment and were affected during maturation by disparate seafloor environmental conditions. The abundance of *M. mosesii* in both groups of environments also suggests that this species implemented during its growing cyst phase a successful adaptive mechanism to oxygen-depleted conditions that might involve anaerobic metabolic interchange with the surrounding media and high level of tolerance to stressors.

The multi-analytical analysis employed in this study, which involves (1) biometry of microphytoplankton fossils, (2) TOFSIMS imaging and surface analysis of selected specimens and (3) host rock geochemistry, has resulted useful for unravelling the interrelation of metabolically active cysts with its surrounding seafloor environment. The causal relationship established between cyst size and environmental conditions allows the biometry of *M. mosesii* to be viewed as a promising prospective vector for massive sulphide and stockwork-type deposits in the IPB.

Supplementary material. For supplementary material/s referred to in this article, please visit <https://doi.org/10.1017/S0016756825100034>

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