

Improvement of mechanical and water absorption properties of plant protein based bioplastics

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1 **Abstract**

2 Bioplastics deriving from plant proteins are becoming an increasingly popular source of
3 raw material for plastic products since they are not only biodegradable but renewable
4 resources. However, these bioplastics require improved mechanical and water
5 absorption properties to be suitable for many applications, such as packaging. For this
6 reason, this study considers potato and rice proteins as a new source for the manufacture
7 of bioplastics. The proteins were mixed with different glycerol concentrations followed
8 by thermomoulding at temperatures from 60 to 180 °C. The resulting bioplastic is
9 characterized in terms of thermo-mechanical properties, water absorption and molecular
10 weight distribution. Compared to well-known wheat gluten, these bioplastics required
11 higher temperatures for their thermomoulding. However, both of them were more
12 structured materials and exhibited less water absorption (e.g. as low as 9 wt.%) than
13 those obtained for wheat gluten blend. Potato protein-based bioplastics showed complex
14 modulus values comparable to synthetic polymers such as Low Density Polyethylene
15 (LDPE).

16 **Keywords:** Rice protein; potato protein; bioplastics; rheology; water absorption;
17 molecular weight properties.

18 **1. Introduction**

19 Bioplastics formulated with plant proteins are becoming an increasingly popular source
20 of raw material for plastic products since they are not only biodegradable but they are
21 also made from renewable resources such as polysaccharides, lipids and proteins
22 (Mooney, 2009; Song & Zheng, 2008; Phillips, A. L., 2008, Wirsenius, Azar &
23 Berndes, 2010). Currently, many bioplastics are still in the developmental stage, but
24 important applications are beginning to emerge in the areas of packaging, food
25 production and medicine (Mooney, 2009; Chen & Tan, 2006; Van de Velde & Kiekens,
26 2002). Some bioplastics can even directly replace synthetically derived materials in
27 traditional applications, whereas others possess unique properties that could open up a
28 range of new commercial opportunities.

29 Their suitability for many applications such as biodegradable films, food packaging or
30 plastic stuffs will depend on material mechanical and water absorption properties.
31 Wheat gluten is a good example of a protein which has been investigated for its
32 thermoplastic properties (Zhong & Yuan, 2013; Jerez et al., 2005a). However, this
33 protein is also highly hygroscopic, which renders it inappropriate for this type of
34 application (Gomez-Martinez et al., 2009; Jerez et al., 2005a). In addition, a critical
35 reason to take into consideration is the potential consumers with celiac disease which
36 comprises approximately 1% of the worldwide population (Renzetti et al., 2012). The
37 use of gluten based bioplastics in food packaging might be risky or even dangerous to
38 their health; the gluten would be in direct contact with food which is a big
39 inconvenience in this case. For this reason, other gluten free alternatives as raw
40 materials should be considered for the manufacturing of bioplastics. In this regard,

41 potato and rice isolates, vegetable proteins that are by-products of the starch industry,
42 are naturally hypoallergenic, meaning that it is entirely free from any gluten.

43 Potato protein has a high content of amino acids with hydrophobic functional groups
44 and its lysine content is much higher than that of other types of protein (Waglay;
45 Karboune & Alli, 2014; Refstie & Tiekstra, 2003). Potato protein can be used as an
46 additive of the feed ingredients, in application of health food, in many food products as
47 emulsifiers and emulsion stabilizers (Waglay; Karboune & Alli, 2014; Singh & Kaur,
48 2009; Van Koningsveld et al., 2006) and can also be used as nitrogen sources for
49 industrial fermentation. Rice protein is high in the sulfur-containing amino acids,
50 cysteine and methionine. This protein could be extracted from rice bran which is an
51 inexpensive, underutilised milling co-product of rough rice (Tang, et al., 2003).

52 Despite the broad available literature about protein based bioplastics, the use of potato
53 or rice proteins as raw materials for manufacture of bioplastics by a thermo-mechanical
54 procedure has so far little studied (Rattanatham et al., 2011; Felix et al., 2016). The
55 purpose of the present study was to evaluate the potential of both proteins isolates as a
56 new source for the production of bioplastics. These bioplastics were obtained by
57 thermomechanical processing that involves a mixing stage followed by
58 thermomoulding, as previously described elsewhere (Gomez-Martinez et al., 2009; Sun,
59 Song & Zheng, 2008; Jerez et al., 2005a; Mangavel et al., 2004; Pommet et al., 2003).
60 The effect of glycerol concentration used as plasticizer, and thermoulding conditions
61 (between 60 to 180°C) on both rice and potato protein based bioplastics was studied.
62 The resulting materials were characterized in terms of thermo-mechanical properties,
63 water absorption and molecular weight distribution.

64 **2. Materials and methods**

65 The bioplastics prepared in this work were composed of rice (R), potato proteins (P)
66 and, as a reference protein, wheat gluten isolates (WG) and glycerol (G) as plasticizer.
67 The proteins were provided by Ferrer Alimentación S.A. (Spain). The composition of
68 rice isolate (R) was about 79 wt.% protein, 5 wt.% lipids, 16 wt.% carbohydrates and
69 2.0 wt.% ashes. Its moisture content was 12 wt.% on dry basis. Potato isolate (P) is
70 mainly composed by 82 wt.% protein, 1.0 wt.% lipids and 0.5 wt. ashes. Its moisture
71 content was 10 wt.% on dry basis. Wheat gluten isolate composition was about 83 wt.%
72 protein, 3 wt.% lipids, 10 wt.% carbohydrates and 1.0 wt.% ashes. Its moisture content
73 was 8 wt.% on dry basis. Glycerol (G) was provided by Prolabo S.A. (Spain).

74 Bioplastic compounds were mixed in a Polylab torque-rheometer equipped with a
75 Rheomix 600p kneading tool (Thermo-Haake GmbH, Germany). This device recorded
76 evolution of temperature and torque along glycerol-protein isolate mixing. Neither
77 heating nor cooling was supplied to the kneading chamber (69 cm³ volume, filled with
78 56g of sample to 85% of its full capacity) during compounding. The process consisted
79 in mixing protein (fine powder) with plasticizer (liquid), by means of two rollers (37.5
80 mm diameter and 47 mm length) counter-rotating at 50 rpm. The mixing time t_{mix} was
81 stopped at 15 min. Subsequently, the resulting blends were compression-moulded into
82 rectangular specimens (50 mm length, 10 mm width, 3 mm thick) by applying a gauge
83 pressure of 100 bar for 10 min at six different temperatures, 90, 100, 120, 140, 160 and
84 180°C (Jerez et al., 2005a,b). These temperatures were above the highest temperature
85 reached during the mixing process. Finally, specimens were allowed to cool down to
86 room temperature inside the hot-plate press before removing from the mould.
87 Afterwards, bioplastics were stored at 53% relative humidity (RH) before testing. Two

88 formulations were studied, containing 33 and 43wt.% glycerol. For sake of simplicity,
89 samples will be identified by the protein source and plasticizer concentration, e.g. potato
90 protein based bioplastics containing 33% and 43% will be referred to as P33 and P43,
91 respectively. Similarly, samples R33, R43, WG 33 and W43 will identify rice and
92 gluten based bioplastics.

93 Dynamic Mechanical Thermal Analysis (DMTA) experiments were performed with a
94 Seiko DMS 6100 (Seiko Instruments, Japan), using 50x10x3 mm³ samples in double
95 cantilever bending mode, according to the ASTM standard method D5023–01 (ASTM,
96 2001b). Storage modulus E' (elastic response) and loss modulus E'' (viscous response)
97 were determined by this technique as a function of temperature. From these data, the
98 complex modulus (E*) was calculated as $|E^*|^2 = |E'|^2 + |E''|^2$. Temperature sweeps were
99 carried out at a constant frequency of 1 Hz, being the applied strain always below
100 0.025% to assure that all tests were within linear viscoelasticity (LVE) region. A
101 heating ramp of 2 °C/min was set between 30 and 180 °C. As a reference, low density
102 polyethylene (LDPE) was measured for comparison with bioplastics.

103 Water absorption measurements were based on ASTM standard method D570-98
104 (ASTM, 2001). Bioplastic samples were dried in an oven, at 50°C for 24 h, and cooled
105 down in a desiccator before weighing (W₀). 50mg of every sample were submerged in
106 50 ml distilled water for a constant period of 24 h. Water on the surface of the samples
107 was removed, and the samples were weighed again (W₁). The remaining water,
108 containing glycerol and soluble proteins, was dried in order to quantify the weight of
109 soluble matter (W_{sol}). The water absorption (Ab) was calculated as follows:

$$110 \quad Ab = \frac{(W_1 - W_0 + W_{sol})}{W_0} \times 100 \quad [1]$$

111 The molecular weight of wheat gluten, rice and potato protein bioplastics was
112 performed by sodium dodecyl sulphate-polyacrylamide gel electrophoresis (SDS
113 PAGE) under reducing conditions as described by Da Silva and Taylor (2004), but
114 using a 4-12% acrylamide gradient gel prepared as described by Byaryhanga et al.
115 (2005). The reducing agent used was 2-Mercaptoethanol, which further denatures the
116 proteins by reducing disulfide linkages, thus overcoming some forms of tertiary protein
117 folding, and breaking up quaternary protein structure (oligomeric subunits).

118

119 At least three replicates (n) were carried out for each test. Standard deviations were
120 calculated for some selected parameters from water uptake capacity and DMTA. The
121 data were presented as average (\bar{x}) \pm standard deviation ($SD = [\sum_n (x_n - \bar{x})^2 / (n -$
122 $1)]^{1/2}$).

123

124 **3. Results and discussion**

125 3.1 Effect of mixing process

126 The mixing process is an important step during manufacturing of protein based
127 bioplastics, in which protein and plasticizer have to be efficiently blended to obtain a
128 material with suitable mechanical properties (or, alternatively, rheology) at the end of
129 the process. The torque and temperature profiles recorded during mixing process of rice
130 (R33, R43) and potato (P33, P43) proteins with 33 and 43 wt.% glycerol are shown in
131 Figure 1. A decrease in torque down to zero was found for all rice protein blends (R33,
132 R43), which became even more evident as the glycerol content increased. On the other
133 hand, the evolution of torque for potato protein blends (P33, P43) shows that the torque

134 values were slightly higher than those recorded for the rice protein blends (Figure 1 A).
135 Furthermore, the temperature variation during the mixing for these samples did not
136 register a continuous increase, as found for other proteins such as wheat gluten (Jerez et
137 al., 2005a), remaining close to 19°C for rice protein blends (R33, R43) and below 22 °C
138 for P43 (Figure 1 B). Similarly, a slight increase in temperature of up to 36°C is
139 observed for a lower glycerol concentration in the potato based blend P33. In any case,
140 this temperature can be considered very low when compared to the temperature reached
141 for wheat gluten samples (Jerez et al., 2005a, Gomez-Martinez et al., 2009), which
142 might suggest a low thermal dissipation of energy and small extent of microstructural
143 changes induced in the protein molecules during mixing, probably due to poor protein-
144 glycerol interactions. This is probably associated with the low molecular weight of both
145 proteins as compared to, for example, wheat gluten (Oszvald et al., 2008). Such a lack
146 of interactions between protein and plasticizer, related to the gentle shear and
147 temperature conditions, allowed the mixing process to be stopped, after 15 min, while
148 the plasticizer-protein dough still showed a suitable rheology (Figure 1A).
149 Subsequently, it was necessary to apply a post-thermal treatment at higher temperature
150 and pressure, in order to improve the interactions between the protein and plasticizer.
151 Therefore, the low mixing temperatures recorded would avoid denaturation or
152 modification of the protein, reported to be close to 80°C by Gorinstein et al. (1996) for
153 rice protein, which in some cases could affect further material processing. Similarly,

154 thermal denaturation (unfolding) of native potato protein is reported between 55-75 °C
155 (Van Koningsveld, 2001), temperature range considerably higher than the temperature
156 recorded during mixing (36 °C). In both cases, the lack of protein denaturation led to a
157 low viscous (deduced from low mixing torque values measured) dough, adequate for
158 further thermo-mechanical processing.

159

160 3.2 Effect of thermo-moulding process

161 The evolution of the complex modulus and $\tan \delta$ with temperature as a function of the
162 thermosetting temperature for rice protein based bioplastic with 33 wt.% glycerol is
163 shown in Figure 2. A decrease in the complex modulus (E^*) with temperature is
164 observed. First, E^* underwent a change in the slope, which occurs at a range of
165 temperature between 74 and 106 °C (Figure 2A). After this, samples exhibited different
166 behaviour depending on the thermosetting temperature. In this sense, a plateau region in
167 E^* was found for samples thermoset below 140 °C, and this event tended to appear at
168 higher temperatures as the thermosetting temperature increased. Conversely, this
169 plateau region was less evident as thermosetting temperature increased up to 160 °C,
170 and at the end of the plateau the complex modulus decreased. On the other hand, for
171 samples thermoset at 180 °C, there was no plateau region, but a continuous decrease in
172 the complex modulus was observed in the temperature range studied. The evolution of
173 $\tan \delta$ with temperature for rice protein based bioplastic with 33 wt.% glycerol at

174 different thermosetting temperatures shows a maximum in $\tan \delta$ at a temperature close
175 to 57 °C for sample thermoset at 90 °C, and 71 °C for sample thermoset at 180 °C
176 (Figure 2B). This maximum is located at higher temperature as thermosetting
177 temperature increased. Also, a decrease in $\tan \delta$ values is observed as thermosetting
178 temperature increased for samples thermoset below 160 °C above which a significant
179 increase in $\tan \delta$ values is reached. In addition, a minimum in $\tan \delta$ at a temperature
180 close to 160 °C for sample thermoset at 90 °C, is found. This minimum appeared at
181 lower temperature as thermosetting temperature increased and did not show up for
182 sample thermoset at 180 °C. Instead, this sample showed a flattening between 137 to
183 160 °C, after which $\tan \delta$ curve decrease.

184 The influence of rice protein on the development of the bioplastic network structure was
185 compared to the structure development in wheat gluten bioplastic with 33 wt.% glycerol
186 and thermoset at 120 °C, as shown in Figure 2. Wheat gluten plasticized with glycerol
187 (henceforth WG/G) was selected as the benchmark because its viscoelastic properties
188 have been widely studied. According to Jerez et al. (2005a), the evolution of the
189 viscoelastic functions for a WG/G sample showed three regions as temperature
190 increased. First, E^* decreased to a plateau region (from 20 to 80 °C) and a maximum in
191 $\tan \delta$ around 30 °C. Second, a plateau region appeared from 100 to 160 °C. Finally,
192 above 160 °C, a decrease in the E^* was observed and a minimum in $\tan \delta$ occurs above
193 125 °C. The rice protein samples displayed E^* values greater than the benchmarked

194 sample up to the first region (from 20 to 80 °C), but beyond 80 °C, the reference sample
195 exhibited about the same value at the plateau region. The maximum peak in loss tangent
196 for the rice protein samples was reached at higher temperature (around 50 °C) than the
197 reference. Although the minimum in loss tangent was at about the same temperature as
198 compared with the benchmarked sample, the values in $\tan \delta$ were significantly higher
199 for rice protein samples than for the WG/G sample.

200 Figure 3 shows the changes in complex modulus and loss tangent with temperature for
201 rice protein based bioplastic with 43 wt.% glycerol, submitted to different
202 thermomoulding temperatures. The complex modulus of the rice protein bioplastics
203 decreased with temperature in the same way as rice protein blends with 33 wt.%
204 glycerol, and also exhibited a change in the slope in E^* , which occurred at temperatures
205 around 100 °C. Moreover, after the change in the slope in the complex modulus,
206 samples thermoset at 120 and 180 °C tended to display a plateau region, whereas a
207 gradual decrease in the slope of the complex modulus was observed for the sample
208 thermoset at 160 °C. A maximum of loss tangent at lower temperatures appeared as the
209 thermosetting temperature increased. The samples also became more elastic (lower loss
210 tangent) as the thermosetting temperature increased, except for the sample thermoset at
211 180 °C (Figure 3 B). An increase in the complex modulus, between 30 and 75 °C, was
212 observed as the thermosetting temperature increased (from 90 to 140 °C). However, an
213 increase in the thermomoulding temperature from 160 to 180 °C led to a decrease in E^* .

214 Therefore, the viscoelastic properties of rice-based bioplastics were affected by glycerol
215 content and thermomoulding temperature treatment. We identified two groups of
216 behaviour, a first group with rice protein samples thermoset at temperature below 140

217 °C and the second one with samples thermoset at temperature above 140 °C. For
218 samples in the first group, it was observed that an increase in glycerol concentration led
219 to a decrease in E^* and glass transition temperature (calculated from the loss tangent
220 peak), although the values at the maximum peak in $\tan \delta$ were not affected. At the same
221 time, an increase in the thermomoulding temperature led to a slight increase in E^* ,
222 which was most evident in the sample with the highest glycerol content. In addition, the
223 glass transition temperature occurred at a range of temperatures between 60–70 °C,
224 while a slight decrease in the values at the maximum peak in $\tan \delta$ was observed. The
225 rice protein samples for the second group, by contrast, showed a significant decrease in
226 E^* , coupled with an almost constant glass transition temperature and a slight increase in
227 the values at the maximum peak in $\tan \delta$. The complex modulus and glass transition
228 temperatures for these samples did not vary with glycerol concentration and
229 thermomoulding temperature.

230 This behaviour could suggest an optimum temperature interval for the rearrangement
231 and alignment of the protein network, in this way promoting the protein-plasticizer
232 interaction and giving more flexibility to the material. According to Ellepola & Ma
233 (2006) rice protein has relatively high thermal stability indicating that it could retain its
234 functionality during the thermomoulding process. These samples also showed a higher
235 viscoelastic modulus than wheat gluten based bioplastic but almost one decade lower
236 than those found for LDPE (Figure 2 and 3).

237 Figure 4 shows the evolution of the complex modulus and $\tan \delta$ with temperature as a
238 function of the thermosetting temperature for potato protein based bioplastic with 33
239 wt.% glycerol. The complex modulus of the potato protein based bioplastic decreased
240 with temperature as expected. Moreover, a change in the slope in E^* was observed. This

241 event occurred within a temperature range of 102-109 °C for samples thermoset at
242 temperatures higher than 120 °C, whilst a change in the slope in E^* at 75 °C was
243 observed for the sample thermoset at 90 °C. There were slight increases in E^* values in
244 the glassy region for samples thermoset below 140 °C after which E^* slightly decreased.
245 The evolution of $\tan \delta$ with temperature for potato protein based bioplastic with 33 wt.%
246 glycerol did not show a minimum value in the loss tangent but a protuberant peak which
247 appeared at a temperature of 75 °C for sample thermoset at 90 °C, and at c.a. 85 °C for
248 sample thermoset at 180 °C. This sample also showed higher values in $\tan \delta$ than other
249 thermosetting temperatures. Non-significant variation in $\tan \delta$ values for samples
250 thermoset below 160 °C was observed.

251 The influence of potato protein on the development of the bioplastic network structure
252 was compared to the structure developed in wheat gluten bioplastic with 33 wt.%
253 glycerol after thermosetting at 120 °C, as shown in Figure 4. The benchmark (i.e. the
254 wheat gluten sample) showed E^* values lower than the potato protein samples across
255 almost the whole range of temperatures studied. These values are more than an order of
256 magnitude higher for potato protein bioplastics than for wheat gluten at low
257 temperature, close to the glassy region (close to 10^9 Pa). In addition, the maximum in
258 loss tangent for the potato protein samples always appeared at a higher temperature (90
259 °C) than the benchmark (30 °C).

260 Figure 5 shows the evolution of complex modulus and loss tangent with temperature for
261 potato-protein based bioplastics with 43 wt.% glycerol. The complex modulus of the
262 potato protein bioplastic decreased with temperature and showed a change in the slope
263 in E^* at a temperature range of 108 to 117 °C. A gradual decrease in the slope of the
264 complex modulus was also observed. In addition, an increase in thermosetting

265 temperature first led to a slight increase in complex modulus values in the glassy region
266 although it then led to a decrease at $T > 140$ °C. A maximum peak in $\tan \delta$ around 80°C
267 for samples with 43 wt.% was found, excepting the sample thermoset at 90 °C (with a
268 peak in $\tan \delta$ at 96°C).

269 The viscoelastic properties of potato protein-based bioplastic did not seem to be too
270 affected by plasticizer concentration or thermosetting treatment as compared to wheat
271 gluten and rice protein bioplastics. The potato protein-based bioplastics had higher
272 complex modulus value (close to 10^9 Pa) and similar to those found for LDPE (Figure 4
273 and 5). Also, $\tan \delta$ peaks for potato protein samples were located at higher temperature
274 than the maximum reached for wheat gluten-based bioplastic and rice systems,
275 indicating that the potato protein blend is more resistant to high temperatures. This
276 behaviour is indicative of a more structured material, with a higher degree of
277 crosslinking.

278 3.3 Characterization of molecular weight distribution

279 In the manufacture of protein-based bioplastics, it is important to understand the
280 composition of the protein and its characteristics, and also how the protein might behave
281 under processing conditions. Understanding these factors is a means of predicting and
282 accounting for possible interactions or reactions between proteins, and the effect of the
283 plasticizers and other components of bioplastics.

284 The molecular weight of wheat gluten, rice and potato protein bioplastics was
285 performed by sodium dodecyl sulphate-polyacrylamide gel electrophoresis (SDS
286 PAGE) under reducing conditions (Figure 6). The results obtained were compared with
287 a protein standard consisting of 12 protein bands in the range of 2.5 – 200 kDa. This

288 reference (Lane 1 in Figure 6) allows accurate molecular weight estimation of the
289 protein sample over a wide molecular weight range.

290 SDS PAGE under reducing conditions showed that the well-known gluten protein
291 bioplastic (Lane 4 in Figure 6) contained bands at 36, 45 and 52 kDa, which
292 corresponded to the alpha, beta and gamma-gliadins, respectively. This range of
293 molecular weight could be also related to the low molecular weight of glutenins (LMW-
294 GS, 20-45 kDa) (Kasarda et al., 1983), which, in our case, might be overlapping. Both
295 proteins tend to be rich in asparagine, glutamine, arginine or proline, serine, methionine
296 and isoleucine (Masci et al., 1995; Lew, Kuzmicky & Kasarda, 1992).

297 In rice protein-based bioplastic (Lane 3 in Figure 6), one large band at 14 kDa can be
298 distinguished. Ogawa et al. (1987) found that rice protein consists of three polypeptide
299 subunits with apparent molecular weight distributions of 10, 13 and 16 kDa. The reason
300 why three bands cannot be found in this sample may be that they are so close that they
301 could not be distinguished from each other. Previous studies have mentioned that 13 and
302 16 kDa are cysteine rich (the sulphur amino acid), which means that this protein might
303 tend to interact and aggregate under thermal conditions (Hibino et al., 1989; Ogawa et
304 al., 1987). Therefore, these three polypeptide subunits might be responsible for the high
305 modulus exhibited in viscoelastic behaviour of rice protein based bioplastics. It is
306 known in the case of wheat gluten based bioplastics that thermomoulding treatment
307 leads to protein aggregation which results in an increase in E^* (Gómez-Martinez et al.,
308 2009). By contrast, in the case of rice protein, E^* decreased for blends thermoset above
309 140 °C as is observed in Figure 2 and 3. It seems that the high temperatures during
310 thermomoulding might help to improve the protein-plasticizer interaction, giving more
311 flexibility to the material (Ellepola & Ma, 2006). Moreover, rice glutelin is extremely

312 insoluble in water because of its hydrophobic, hydrogen and disulfide bonding
313 (Hamada, 1996; Juliano, 1985). In addition to the rice protein sample characterization,
314 this sample exhibited one band at 21 kDa close to the band (25 kDa) cited by many
315 authors for rice albumins which have a wide range of molecular weights (Houston &
316 Mohammed 1970). However, according to Juliano (1972), the major component of the
317 glutelin is located at 18–20 kDa and thus close to the band we found.

318 The potato protein-based bioplastic (Lane 2 in Figure 6) contained a group of proteins at
319 6, 16, 21 and 45 kDa. The first three bands mentioned might be identified by protease
320 inhibitors. A characteristic of this protein is that it is small, cysteine rich (comprising a
321 large number of disulphide bridges) and heat-resistant (Pouvreau et al., 2005a,b; Van
322 Koningsveld, 2001). This fact might promote the aggregation of the protein under
323 thermal conditions and may play an important role in the viscoelastic behaviour
324 properties of this material. Therefore, this factor may explain the high values of E^* and
325 $\tan \delta$ peaks located at high temperature (see Figure 2 and 3). Also, thanks to this
326 individual protein the potato protein blend was more resistant to high temperatures than
327 wheat gluten bioplastics and rice protein systems. According to Pouvreau et al.
328 (2005a,b), protease inhibitors is an individual protein with heat-resistant properties
329 which might explain the low sensitivity of these blends to be affected by the
330 thermosetting temperature as compared to wheat gluten and rice protein samples. The
331 45 kDa band might be identified as patatin which is a hydrophobic protein (Park et al.,
332 1983). In addition, two weak bands at high molecular weights 54 and 66 kDa were
333 found. These types of oligomers represent about 12 wt.% of the protein present, such as
334 lectin (Pouvreau et al., 2001; Allen et al., 1996).

335 3.4 Water absorption

336 Previous results have shown the type of protein used and thermomechanical treatment
337 can lead to different thermo-mechanical behaviour and, likewise, different water uptake
338 capabilities should be expected. Water absorption is an important parameter to be
339 controlled when material application or processing is required (Rouilly et al., 2005;
340 Jerez et al., 2005a, 2007). Figure 7 shows the water absorption values for rice and
341 potato protein-based bioplastics manufactured with different glycerol content at
342 different thermosetting temperatures.

343 The observed water absorption values for rice protein based bioplastics with 33 wt%
344 glycerol showed a continuous decrease with increasing thermosetting temperature, from
345 90 to 180°C (Figure 7A). Thus, samples thermoset above 160°C exhibited the lowest
346 water absorption, about 20%. Conversely, at 43 wt% glycerol content, absorption
347 tended to decrease with higher thermosetting temperatures until these reached 140°C.
348 This behaviour agrees with the highest values of E^* for this blend (see Figure 3), after
349 which it increased to around 33 wt.%.

350 Figure 7B shows the water absorption values for potato protein-based bioplastics
351 manufactured at different thermosetting temperatures and different glycerol
352 concentrations. The water absorption values for the samples with 33 wt.% glycerol
353 decrease for samples thermoset below 140 °C which registered the lowest absorption
354 value (9 wt.%), after which sample gave higher values with increased thermosetting
355 temperatures. These results are entirely consistent with the values in the complex
356 modulus observed at these thermosetting temperatures (see Figure 4). By contrast,
357 potato protein-based bioplastics with 43 wt.% glycerol did not show a clear trend with
358 thermosetting temperature. This blend exhibited the minimum value in water absorption
359 with the sample thermoset at 180 °C (13 wt.%).

360 On the whole, water absorption variability among replicates for both proteins decreased
361 as temperature thermosetting increased, coupled to the progressive thermal denaturation
362 of proteins. In any case, the obtained results pointed out that both protein isolates led to
363 materials significantly more hydrophobic than the reference gluten-derived bioplastics,
364 which typically exhibit water absorption values in the range 80-90 wt.% (Gomez-
365 Martinez et al., 2009; Jerez et al., 2007).

366 In general, thermomoulding treatment leads to protein denaturation which results in an
367 increase of hydrophobicity, due to exposure of hydrophobic groups that are folded
368 inside the intact native protein molecule (Mine, 1997), showing that E^* increases as
369 water absorption decreases (Gomez-Martinez et al., 2009; Jerez et al., 2007). By
370 contrast, rice and potato protein samples did not show the same tendency. Both proteins
371 however exhibited water absorption values lower than 50 wt.%, these values are low
372 compared to those obtained for wheat gluten blend (Gomez-Martinez et al., 2009). This
373 behaviour is probably due to the presence of glutelin in the rice protein and the patatin
374 in potato protein, which are not present in wheat gluten (Figure 6-Line 4). They are
375 extremely insoluble in water because of its hydrophobic, hydrogen and disulfide
376 bonding (Oszvald et al., 2008; Hamada, 1996; Juliano, 1985; Park et al., 1983).
377 Therefore, these two polypeptide subunits may play an important role in reducing the
378 water absorption for these two bioplastics. This fact was more evident in the absorption
379 properties of the potato protein which brings the lowest water uptake as well as a
380 narrower range of these values compared to rice protein-based bioplastics.

381 **Conclusions**

382 The resulting blends from potato and rice proteins, as new raw materials to manufacture
383 bioplastics, have exhibited suitable viscoelastic and water absorption properties to be
384 potentially used as food packaging.

385 The mixing process hardly induces protein-plasticizer interactions, being required a
386 post-thermal treatment at higher temperature and pressure to obtain a material with
387 suitable properties. Potato protein-based bioplastics had a complex modulus value
388 similar to those found for LDPE. The viscoelastic properties of rice protein-based
389 bioplastic were affected by glycerol content and thermomoulding temperature treatment.
390 In contrast, potato protein-based bioplastic did not seem to be affected.

391 Both proteins exhibited water absorption values lower than 50 wt.%, although potato
392 protein based bioplastic showed the lowest water absorption (9 wt.%). These values are
393 significantly lower than those obtained for wheat gluten blends. In this regard, glutelin
394 in the rice protein, and the protease inhibitors and patatin in potato protein, may play an
395 important role in both viscoelastic behaviour and water absorption properties of rice and
396 potato protein bioplastics.

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401 **References**

402 Allen, A. K., Bolwell, G. P., Brown, D. S., Sidebottom C., & Slabas, A. R. (1996). Potato lectin:
403 A three domain glycoprotein with novel hydroxyproline-containing sequences and sequence

404 similarities to wheat germ agglutinin. *International Journal of Biochemistry & Cell Biology*, 28,
405 1285-1291.

406 ASTM, (2001a). Standard test method for Plastics: Dynamic Mechanical Properties: In Flexure
407 (Three-Point Bending). Designation D 5023–01. In Annual book of ASTM standards
408 Philadelphia, PA: American Society for Testing and Materials.

409 ASTM, (2001b). Standard test method for Plastics: Water Absorption of Plastics Designation D
410 570 – 98. In Annual book of ASTM standards Philadelphia, PA: American Society for Testing
411 and Materials.

412 Byaruhanga, Y. B., Erasmus, C., & Taylor, J. R. N. (2005). Effect of microwave heating of
413 kafirin on the functional properties of kafirin films. *Cereal Chemistry Journal*, 82, 565-573.

414 Chen, Y., & Tan, H.M. (2006). Crosslinked carboxymethylchitosan-g-poly (acrylic acid)
415 copolymer as a novel superabsorbent polymer. *Carbohydrate Research*, 341, 887-896.

416 Da Silva, L. S., & Taylor, J. R. N. (2004). Sorghum bran as a potential source of kafirin. *Cereal*
417 *Chemistry Journal*, 81, 322-327.

418 Ellepola, S.W., & Ma, C.-Y. (2006). Thermal properties of globulin from rice (*Oryza sativa*)
419 seeds. *Food Research International*, 39 (3), 257-264.

420 Félix, M., Lucio-Villegas, A., Romero, A. & Guerrero, A. (2016). Development of rice protein
421 bio-based plastic materials processed by injection molding. *Industrial Crops and Products*, 79,
422 152-159.

423 Gomez-Martinez, D. P., Partal, P., Martinez, I., & Gallegos, C. (2009). Rheological behaviour
424 and physical properties of controlled-release gluten-based bioplastics. *Bioresource Technology*,
425 100, 1828-1832.

426 Gorinstein, S., Zemser, M., Friedman, M., Rodrigues, W. A., Martins, P. S., Vello, N. A.,
427 Tosello, G. A., & Paredes-López, O. (1996). Physicochemical characterization of the structural
428 stability of some plant globulins. *Food Chemistry*, 56, 131-138.

429 Gujral, H. S., & Rosell, C. M. (2004). Improvement of the bread making quality of rice flour by
430 glucose oxidase. *Food Research International*, 37, 75-81.

431 Hamada, J. S. (1996). Separation and molecular mass distribution of rice proteins by size-
432 exclusion high-performance liquid chromatography in a dissociating buffer. *Journal of*
433 *Chromatography*, 734, 195–203.

434 Hibino, T., Kidzu, K., Masumra, T., Ohtsuki, K., Tanaka, K., Kawabata, M., & Fujii, S. (1989).
435 Amino-acid composition of rice prolamin polypeptides. *Agricultural and Biological Chemistry*,
436 53, 513–518.

437 Houston, D.F. & Mohammed, A. (1970). Purification and partial characterization of a major
438 globulin from rice endosperm *Cereal Chemistry*, 47, 5-12.

439 Jerez, A., Partal, P., Martínez, I., Gallegos, C., & Guerrero, A. (2005a). Rheology and
440 processing of gluten based bioplastics. *Biochemical Engineering Journal*, 26, 131-138.

441 Jerez, A., Partal, P., Martínez, I., Gallegos C., & Guerrero, A. (2005b). Bioplástico y métodos
442 para su preparación. Oficina Española de Patentes y Marcas. Patent, P200501556.

443 Jerez, A., Partal, P., Martínez, I., Gallego, C., & Guerrero, A. (2007). Egg white-based
444 bioplastics developed by thermomechanical processing. *Journal of Food Engineering*, 82, 608 –
445 617.

446 Juliano, B. (1972). The rice caryopsis and its composition. In D. F. Houston (Ed.), *Rice:*
447 *Chemistry and Technology* (pp 16-74). St. Paul, American Association of Cereal Chemists Inc.

448 Juliano, B. (1985). Polysaccharides, proteins and lipids of rice. In: B. O. Juliano (Ed.), *Rice:*
449 *Chemistry and Technology* (pp.59-179). St. Paul, American Association of Cereal Chemists, Inc.

450 Kasarda, D. D., Autran, J. C., Lew, E. J. L., Nimmo, C. C., & Shewry, P. R. (1983). N-terminal
451 amino acid sequences of ω -gliadins and ω -secalins, Implications for the evolution of prolamin
452 genes. *Biochimica et Biophysica Acta*, 747, 138-150.

453 Lew, E. J. L., Kuzmicky, D. D., & Kasarda, D. D. (1992). Characterization of low-molecular-
454 weight glutenin subunits by reversed-phase highperformance liquid chromatography, sodium

455 dodecyl sulfate-polyacrylamide gel electrophoresis, and N-terminal amino acid sequencing.
456 *Cereal Chemistry Journal*, 69, 508–515.

457 Mangavel, C., Rossignol, N., Perronnet, A., Barbot, J., Popineau, Y., & Gueguen, J. (2004).
458 Properties and microstructure of thermo-pressed wheat gluten films: A comparison with cast
459 films. *Biomacromolecules*, 5, 1596–1601.

460 Masci, S., Lew, E. J.-L., Lafiandra, D., Porceddu, E., & Kasarda, D. D. (1995). Characterization
461 of low-molecular-weight glutenin subunits in durum wheat by RP-HPLC and N-terminal
462 sequencing. *Cereal Chemistry*, 72, 100–104.

463 Mine Y. (1997). Effect of dry heat and mild alkaline treatment on functional properties of egg
464 white proteins. *Journal of Agricultural and Food Chemistry*, 45, 2924-2928.

465 Mooney B.P. (2009). The second green revolution? Production of plant-based biodegradable
466 plastics. *Biochemical Journal*, 418, 219–232.

467 Ogawa, M., Kumamaru, T., Satoh, H., Iwata, N., Omura, T., Kasai, Z., & Tanaka, K. (1987).
468 Purification of protein body I of rice seed and its polypeptide composition. *Plant and Cell*
469 *Physiology*, 28, 1517–1527.

470 Oszvald, M., Tömösközi, S., Larroque, O., Keresztényi, E., Tamás, L., & Békés, F. (2008).
471 Characterization of rice storage proteins by SE-HPLC and micro z-arm mixer. *Journal of Cereal*
472 *Science*, 48, 68-76

473 Oszvald, M., Tömösközi, S., Tamás, L., & Békés, F. (2007). Developing method to study the
474 effect of rice proteins for the functional properties. *Acta Alimentaria*, 37, 399-408

475 Park, W. D., Blackwood, C., Mignery, G. A., Hermodson, M. A., & Lister, R. M. (1983).
476 Analysis of the heterogeneity of the 40,000 molecular weight tuber glycoprotein of potatoes by
477 immunological methods and by NH₂-terminal sequence analysis. *Plant Physiol.*, 71, 156–160

478 Phillips, A. L. (2008). Bioplastics Boom. *American Scientist*, 96, 190

479 Pommet, M., Redl, A., Morel, M. H., Domenek, S., & Guilbert, S. (2003). Thermoplastic
480 processing of protein-based bioplastics: chemical engineering aspects of mixing, extrusion and
481 hot molding. *Macromolecular Symposia*, 197, 207–17.

482 Pouvreau, L., Gruppen, H., Piersma, S. R., Van den Broek, L. A. M., Van Koningsveld, G. A.,
483 & Voragen, A. G. J. (2001). Relative abundance and inhibitory distribution of protease
484 inhibitors in potato fruit juice from c.v. Elkana. *Journal of Agricultural and Food Chemistry*,
485 49, 2864–2874.

486 Pouvreau, L., Gruppen, H., Van Koningsveld, G., Van den Broek, L. A. M., & Voragen, A. G.
487 J. (2005a). Conformational stability of the potato serine protease inhibitor group. *Journal of*
488 *Agricultural and Food Chemistry*, 53, 3191–3196.

489 Pouvreau, L., Kroef, T., Gruppen, H., Van Koningsveld, G., Van den Broek, L. A. M., &
490 Voragen, A. G. J. (2005b). Structure and stability of the potato cysteine protease inhibitor group
491 (Cv. Elkana). *Journal of Agricultural and Food Chemistry*, 53, 5739–5746.

492 Rattanatham, P., Kunanopparat, T. & S. Siriwattanayotin, S. (2011). improvement of
493 rheological and functional properties of defatted rice bran protein bioplastic by kraft lignin
494 Addition. *Thai Journal of Agricultural Science*, 44(5), 56-61.

495 Refstie, S., & Tiekstra, H. A. J. (2003). Potato protein concentrate with low content of
496 solanidine glycoalkaloids in diets for Atlantic salmon (*Salmo salar*). *Aquaculture*, 216, 283–298

497 Renzetti, S., Behr, J., Vogel, R. F., Barbiroli, A., Iametti, S., Bonomi, F., & Arendt, E.K.
498 (2012). Transglutaminase treatment of brown rice flour: A chromatographic, electrophoretic and
499 spectroscopic study of protein modifications. *Food Chemistry*, 131, 1076–1085.

500 Rouille, J., Della Valle, G., Lefebvre, J., Sliwinski, E., & VanVliet, T. (2005). Shear and
501 extensional properties of bread doughs affected by their minor components. *Journal of Cereal*
502 *Science*, 42(1), 45-57.

503 Sun, S., Song Y., & Zheng Q. (2008). Thermo-molded wheat gluten plastics plasticized with
504 glycerol: Effect of molding temperature. *Food Hydrocolloids*, 22, 1006–1013

505 Singh, J., & Kaur, L. (2009). Introduction: potato tuber. In Singh, J. Kaur, L. (Eds.). *Advances*
506 *in Potato Chemistry and Technology* (pp. 9-12). Burlington, MA: Elsevier.

507 Sivaramakrishnan, H. P., Senge, B., & Chattopadhyay, P. K. (2004). Rheological properties of
508 rice dough for making rice bread. *Journal of Food Engineering*, *62*, 37-45.

509 Song, Y., & Zheng, Q. (2008). Improved tensile strength of glycerol-plasticized gluten
510 bioplastic containing hydrophobic liquids. *Bioresource Technology*, *99*, 7665-7671.

511 Tang, S., Hettiararchy, N. S., Eswaranandam, S. & Crandall, P. (2003). Protein extraction from
512 heat-stabilized defatted rice bran II. The role of amylase, celluclast, and viscozyme. *Journal of*
513 *Food Science*, *68*, 471–475.

514 Van de Velde, K., & Kiekens, P. (2002). Biopolymers: overview of several properties and
515 consequences on their applications. *Polymer Testing*, *21*, 433–442

516 Van Koningsveld, G.A., Walstra, P., Voragen, A. G. J., Kuijpers, I. J., Van Boekel, M. A. J. S.,
517 & Gruppen, H. (2006). Effects of protein composition and enzymatic activity on formation and
518 properties of potato protein stabilized emulsions. *Journal of Agricultural and Food Chemistry*,
519 *54*, 6419-6427.

520 Van Koningsveld, G. A., Gruppen, H., de Jongh, H. H. J., Wijngaards, G., Van Boekel, M. A. J.
521 S., & Walstra, P. (2001). Effects of pH and heat treatment on the structure and solubility of
522 potato proteins in different preparations. *Journal of Agricultural and Food Chemistry*, *49*,
523 4889–4897.

524 Waglay, A., Karboune S., & Alli, I. (2014). Potato protein isolates: Recovery and
525 characterization of their properties. *Food Chemistry*, *14*, 373-382

526 Wirsenius, S., Azar, C., & Berndes, G. (2010). How much land is needed for global food
527 production under scenarios of dietary changes and livestock productivity increases in 2030?.
528 *Agricultural Systems*, *103*, 621-638.

529 Zhong, N. & Yuan, Q. (2013). Preparation and properties of molded blends of wheat gluten and
530 cationic water-borne polyurethanes. *Journal of Applied Polymer Science*, *128* (1), 460-469.

FIGURES

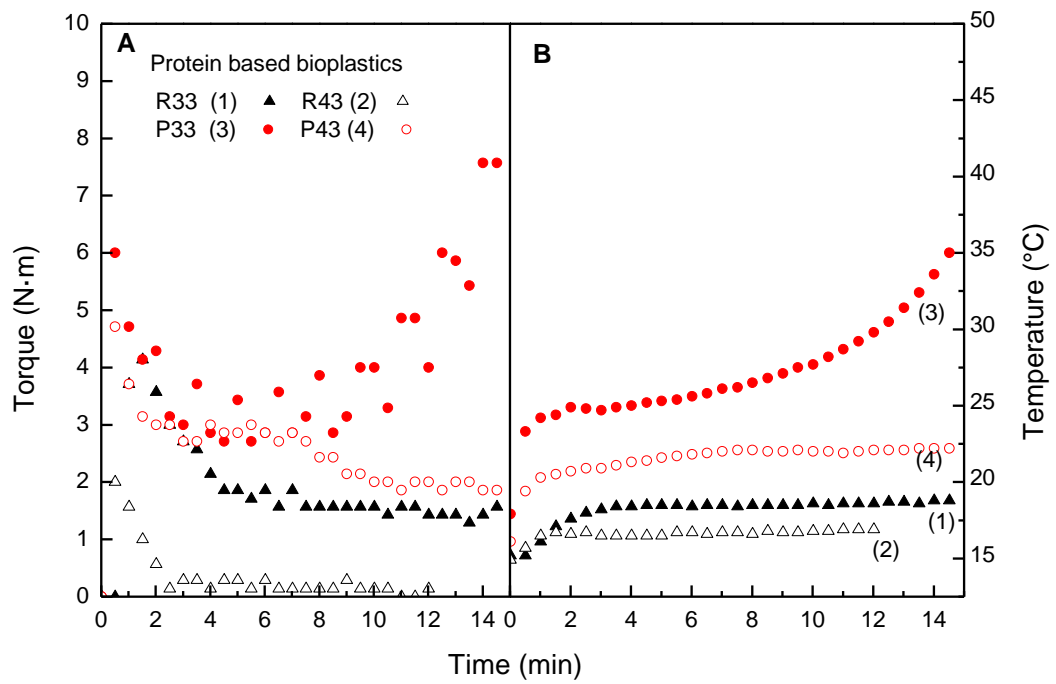


Figure 1. Evolution of torque (A) and temperature (B) during the mixing process for rice and potato protein-based bioplastic with varying glycerol content.

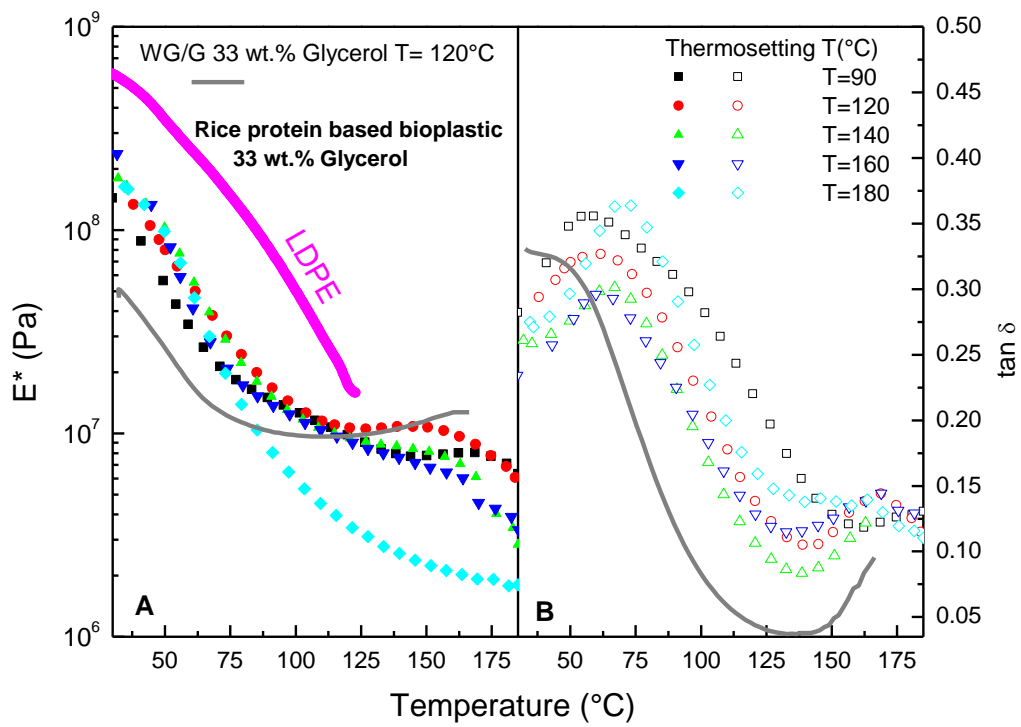


Figure 2. Dynamic mechanical thermal analysis results, complex modulus E^* (A) and $\tan \delta$ (B) for rice protein based bioplastic with 33 wt.% glycerol (R33) at different thermosetting temperatures.

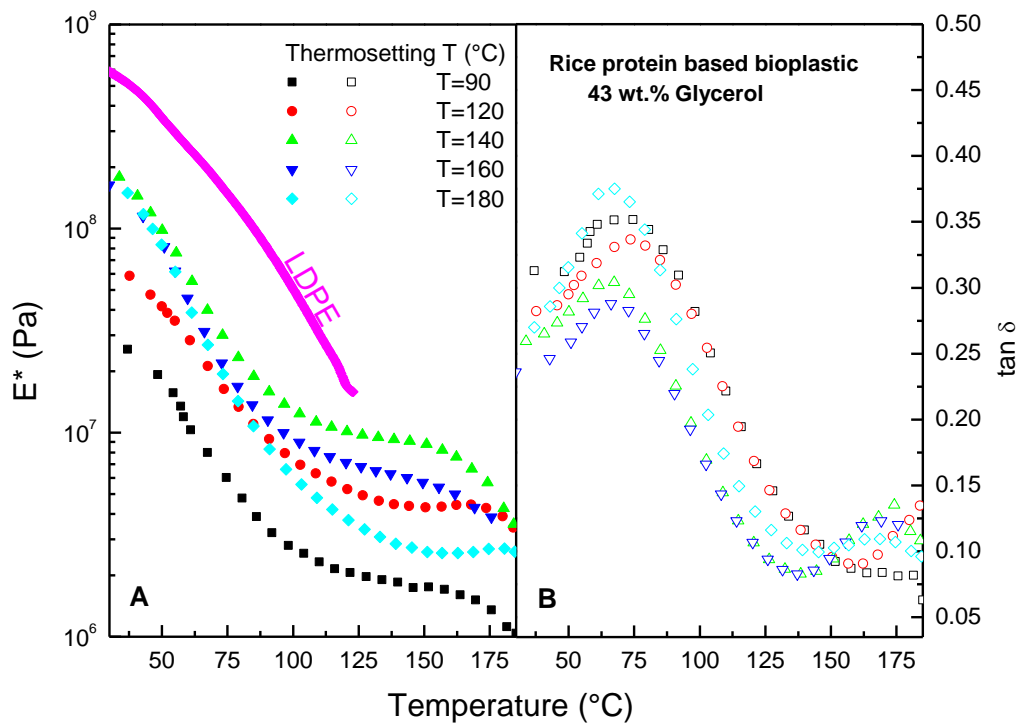


Figure 3. Dynamic mechanical thermal analysis results for the complex modulus E^* (A) and $\tan \delta$ (B) for rice protein based bioplastic with 43 wt.% glycerol (R43) at different thermosetting temperatures.

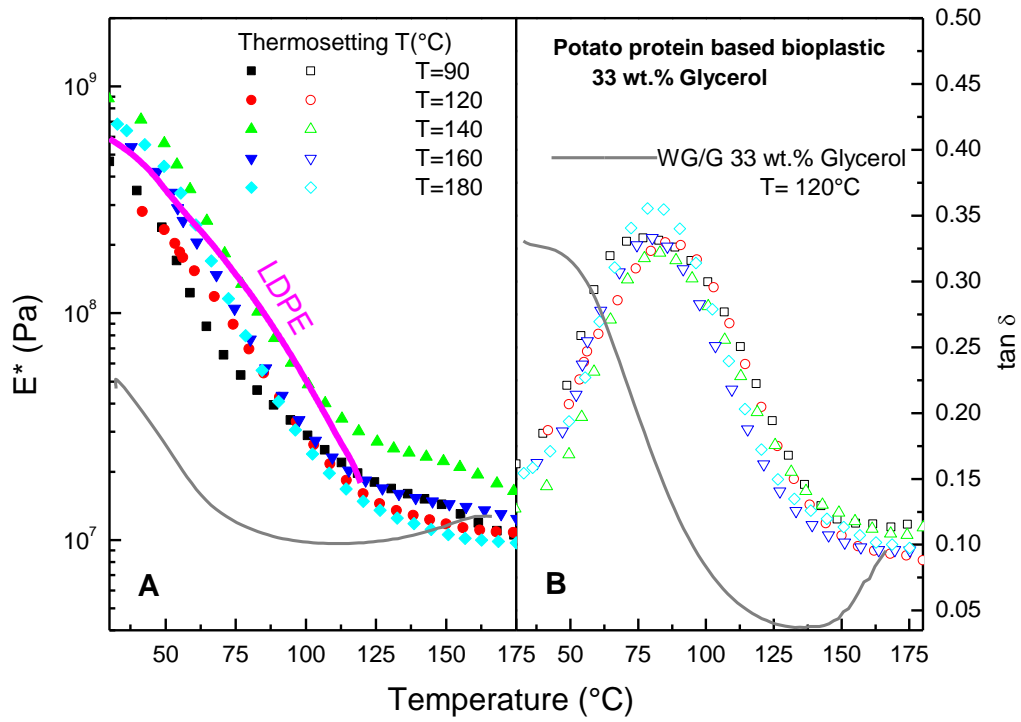


Figure 4. Dynamic mechanical thermal analysis results, complex modulus E^* (A) and $\tan \delta$ (B) for potato protein based bioplastic with 33 wt.% glycerol (P33) at different thermosetting temperatures.

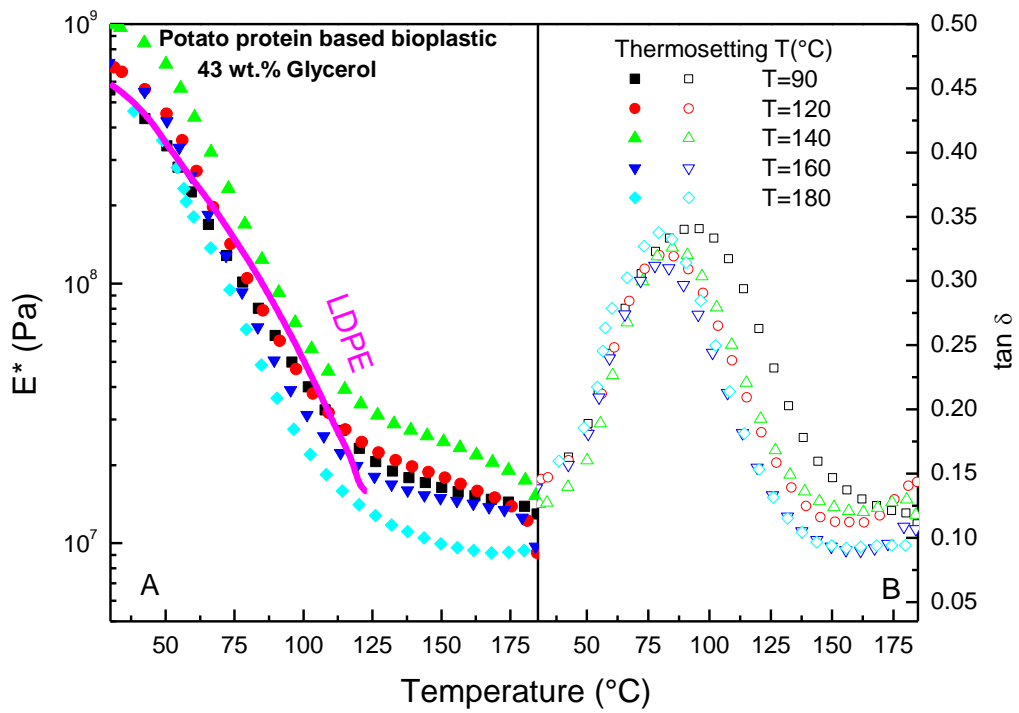


Figure 5. Dynamic mechanical thermal analysis results showing the complex modulus E^* (A) and $\tan \delta$ (B) for potato protein based bioplastic with 43 wt.% glycerol (P43) at different thermosetting temperatures.

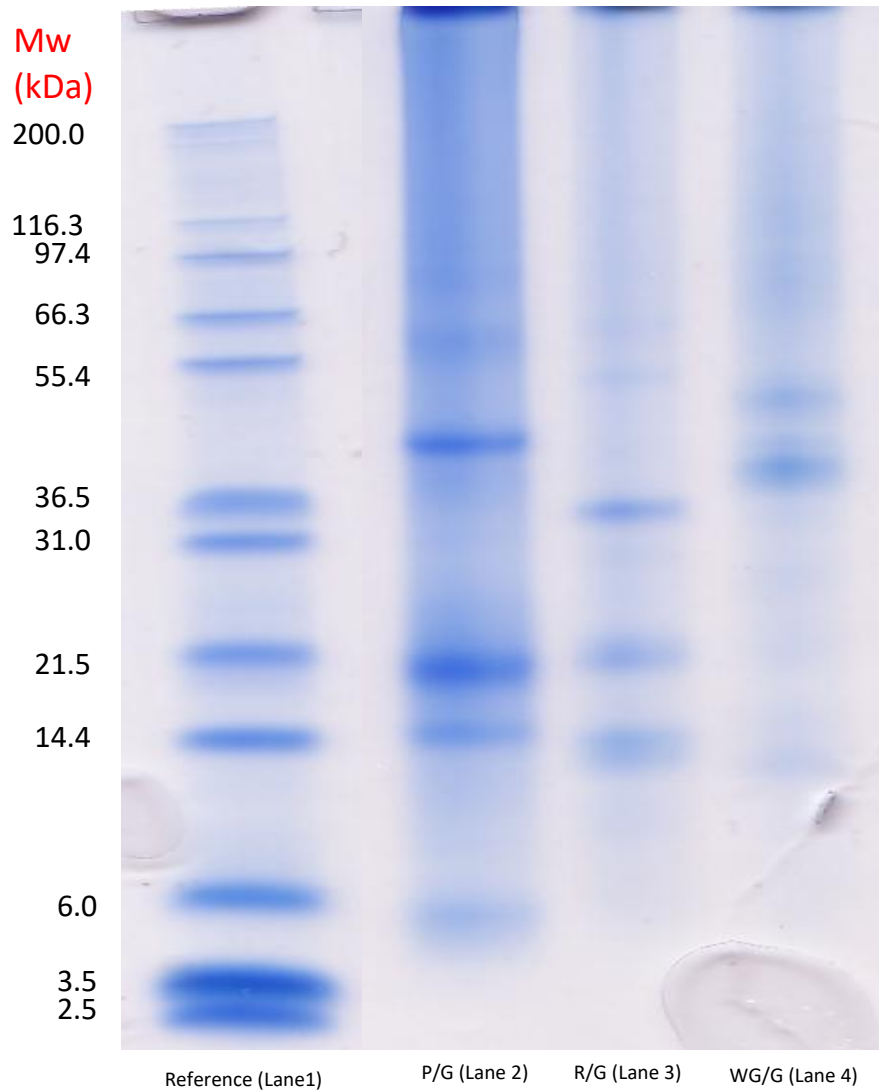


Figure 6. SDS-PAGE for wheat gluten, rice and potato protein based bioplastics, under reducing conditions. Lane 1: reference, lane 2: potato protein/glycerol(P/G), lane 3: rice protein/glycerol(R/G) and lane 4: wheat gluten/glycerol (WG/G) under reducing conditions.

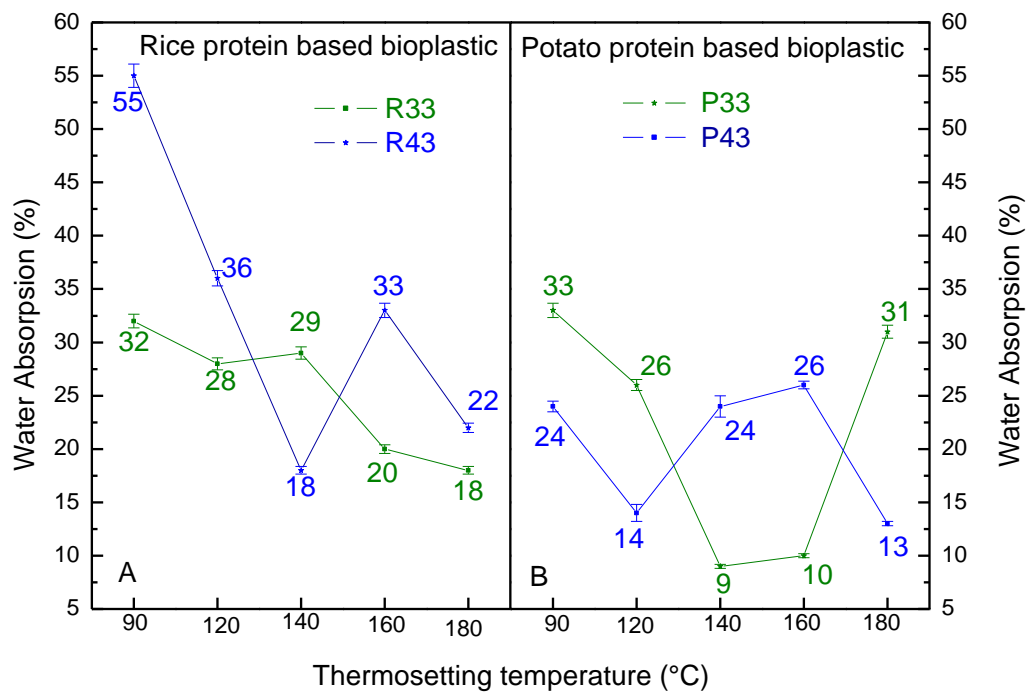


Figure 7. Water absorption for rice and potato protein bioplastics plasticized with different glycerol concentrations, at different thermosetting temperatures